

ARTICLE INFO

Received 02 December 2019

Accepted 13 December 2019

Available online December 2019

STEREOCONTROLLED SYNTHESIS OF FLUORINE-CONTAINING FUNCTIONALIZED β -LACTAM DERIVATIVES THROUGH CROSS-METATHESIS REACTIONS

Attila M. Remete,^{1,2} Zsanett Benke,^{1,2} Loránd Kiss^{1,2*}

¹*Institute of Pharmaceutical Chemistry, University of Szeged, H-6720 Szeged, Eötvös u. 6, Hungary*

²*University of Szeged, Interdisciplinary Excellence Centre, Institute of Pharmaceutical Chemistry, H-6720 Szeged, Eötvös u. 6, Hungary*

E-mail: kiss.lorand@pharm.u-szeged.hu

Abstract: Functionalized β -lactams and their derivatives have generated increasing interest in medicinal chemistry over the last decades. Because of their pharmacological potential the chemistry of fluorinated β -lactams and β -amino acids is considered to be an expanding research field. The stereocontrolled synthesis of various fluorine-containing acyclic β -lactams has been accomplished from some readily available unsaturated bicyclic β -lactam isomers. The synthetic strategy was based on ring-opening metathesis of unsaturated bicyclic azetidiones followed by cross-metathesis with fluorine-containing olefins. The cross-metathesis transformations, performed under various conditions with the aim of studying chemodiscrimination of the olefin bonds, resulted in the corresponding functionalized β -lactam derivatives.

Keywords: β -lactam, ring opening, functionalization, metathesis, selectivity, fluorine

Introduction

Azetidin-2-ones or β -lactams have received considerable attention in organic and medicinal chemistry. These compounds exhibit a wide range of biological activities. β -Lactam antibiotics are amongst the most important antibacterial agents [1, 2]. Other reported activities include protease inhibition, cholesterol absorption inhibition, human fatty acid synthase inhibition, and antitumor properties [3, 4]. β -Lactams are also valuable synthetic intermediates towards heterocycles, β -amino acids, peptides, taxoids, and other compounds [5-8].

Because of the high importance of fluorinated biomolecules in drug design, the introduction of one or more fluorine atoms into an organic molecule has attracted high interest over the past ten

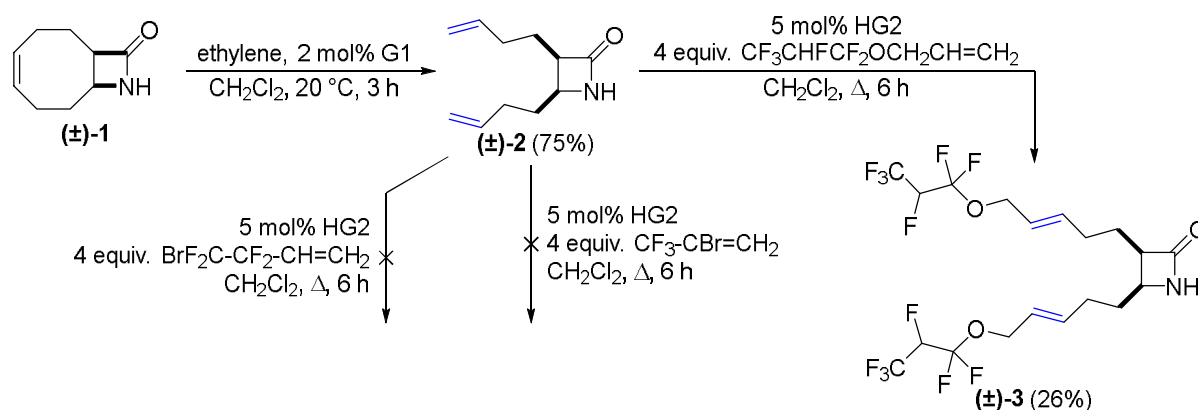
years [9-13]. Thanks to the utility of β -lactams, this interest has also been extended to fluorine-containing azetidin-2-ones. Such compounds are often synthesized by using a fluorinated reactant during heterocyclization [14-19], but late-stage fluorination (fluorine introduction after synthesizing the azetidinone skeleton) can also be applied [20]. β -Lactams expressed their importance in the synthesis of different fluorinated molecules such as sugar- β -amino acid conjugates [15], amido esters [18], aminopropanes and heterocycles [17], as well as taxoids and β -amino acids [20]. Relevant bioactivities were reported for fluoro-taxoids [20] and some *N*-aryl-3,3-difluoroazetidin-2-ones [21].

One of the most important synthetic developments in the last two decades was the emergence of olefin metathesis reactions. This breakthrough was made possible by commercially available, highly active Ru-based catalysts tolerating varied functional groups (common abbreviations: G1 for Grubbs 1st generation catalysts, G2 for Grubbs 2nd generation catalysts, HG1 for Hoveyda-Grubbs 1st generation catalysts, and HG2 for Hoveyda-Grubbs 2nd generation catalysts) [22, 23]. This powerful, new pathway enabled the synthesis of various compounds, which were previously impossible or hard to prepare [24, 25].

Results and Discussion

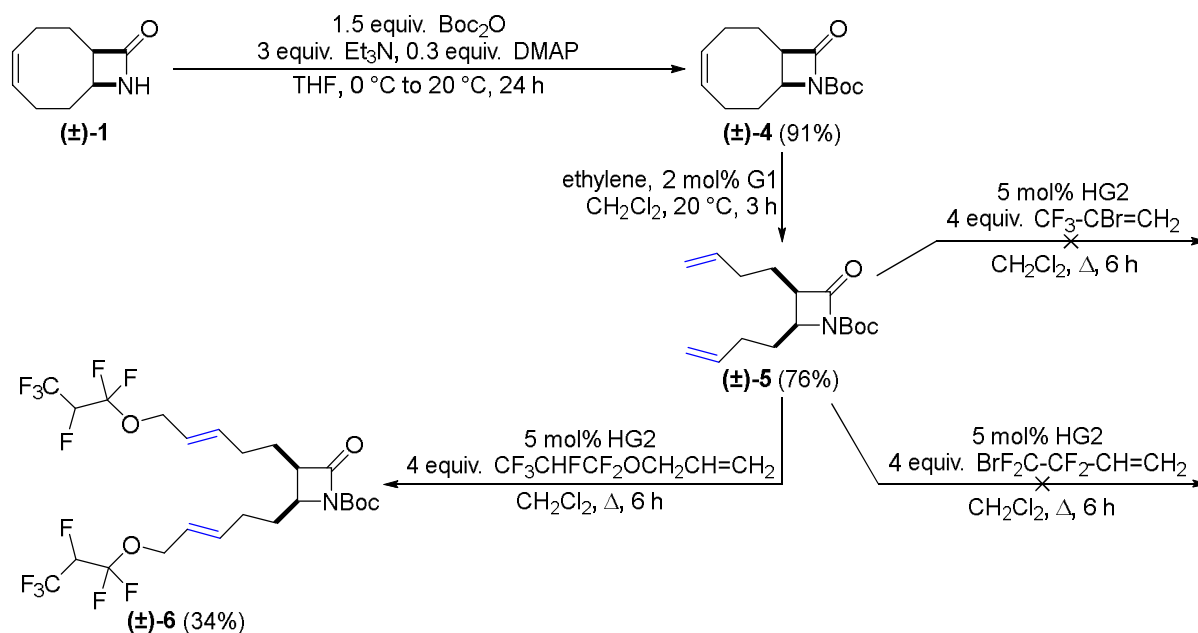
In our earlier recent studies, we have successfully transformed unsaturated bicyclic β -lactams into functionalized monocyclic azetidinones via a ring-opening metathesis/cross metathesis (ROM/CM) sequence [26, 27]. Our current aim was to extend this stereocontrolled pathway to the synthesis of fluorine-containing β -lactams utilizing fluorinated olefins in the CM step.

Our first selected substrate was bicyclic lactam (\pm)-**1** derived from 1,5-cyclooctadiene. Its ring opening with ethylene was accomplished according to a literature method [26]. Functionalization of alkenyl-substituted lactam (\pm)-**2** thus obtained was attempted with three different commercially available highly fluorinated alkenes. Cross metathesis reactions in the presence of the four commercial catalysts (G1, G2, HG1, HG2) with 2-bromo-3,3,3-trifluoro-1-propene and 4-bromo-3,3,4,4-tetrafluoro-1-butene failed, but the reaction with allyl 1,1,2,3,3,3-hexafluoropropyl ether proved to be successful. The best yield of dicoupled product (\pm)-**3** was achieved with catalyst HG2 (Scheme 1).



Scheme 1. Cross-metathesis of lactam **(±)-2** with fluorinated alkenes.

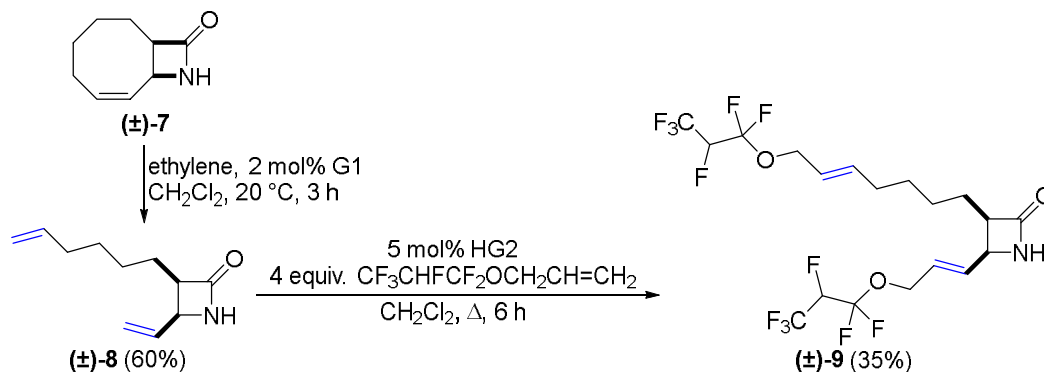
Compound **(±)-4**, our next selected substrate, was obtained by *N*-Boc protection of lactam **(±)-1**. Modification of the reported procedure [28] resulted in an increased product yield of this step. ROM of compound **(±)-4** was performed analogously to ring opening of **(±)-1**. Product **(±)-5** formed in 77% yield was then subjected to CM in the presence of catalyst HG2. Similar to cross metathesis of **(±)-2**, the reaction succeeded with allyl 1,1,2,3,3,3-hexafluoropropyl ether to afford compound **(±)-6**, but failed with 2-bromo-3,3,3-trifluoro-1-propene and 4-bromo-3,3,4,4-tetrafluoro-1-butene (Scheme 2).



Scheme 2. Cross-metathesis of Boc-protected lactam **(±)-5** with fluorinated alkenes.

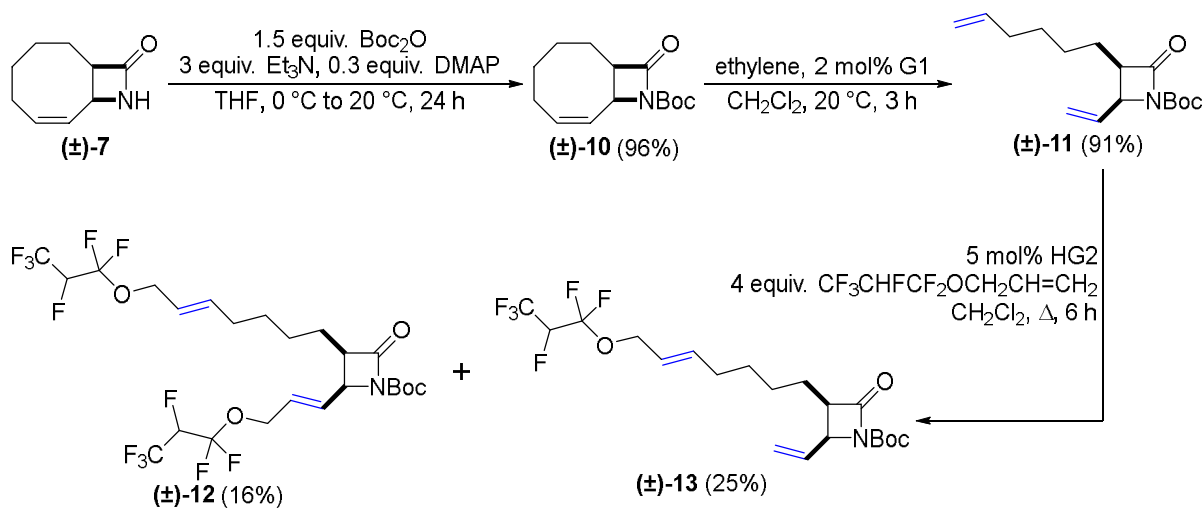
We continued our work with bicyclic lactam **(±)-7** (a regioisomer of lactam **(±)-1**) derived from 1,3-cyclooctadiene. Its ring opening with ethylene was accomplished according to a literature method described earlier [26]. Taking into account the CM reactions on Scheme 1 and Scheme 2,

functionalization of the resulting (\pm)-**8** was attempted only with allyl 1,1,2,3,3,3-hexafluoropropyl ether. The transformation, carried out in the presence of catalyst HG2, gave dicoupled product (\pm)-**9** in a yield of 35% (Scheme 3).



Scheme 3. Cross-metathesis of lactam (\pm)-**8** with fluorinated alkene.

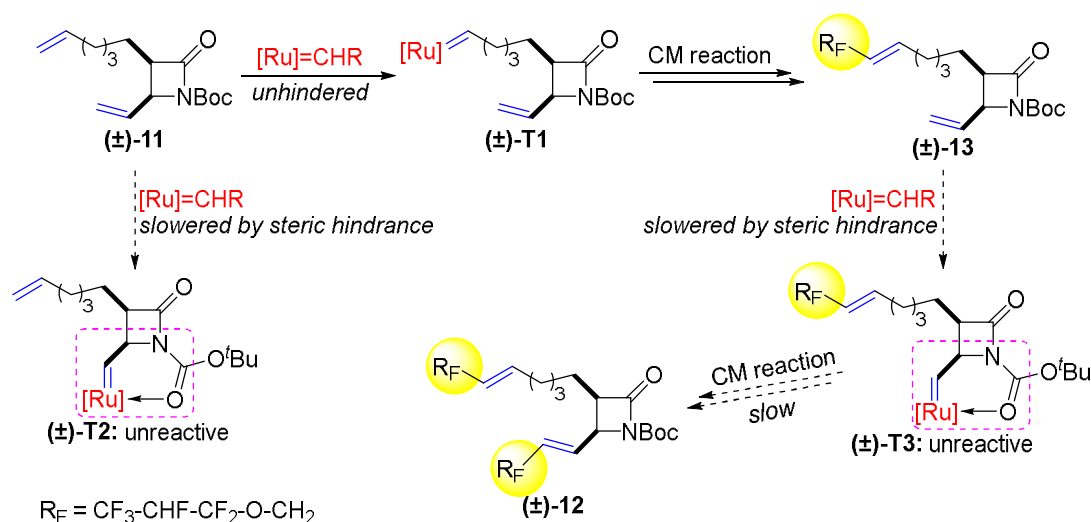
Next, bicyclic lactam (\pm)-**10** (*N*-Boc-protected derivative of (\pm)-**7**) was synthesized. Its reaction with ethylene was performed analogously to ROM of compounds (\pm)-**1** and (\pm)-**4**. Cross metathesis of the obtained (\pm)-**11** with allyl 1,1,2,3,3,3-hexafluoropropyl ether produced not only dicoupled product (\pm)-**12** but the formation of monocoupled product (\pm)-**13** was also observed (Scheme 4).



Scheme 4. Cross-metathesis of lactam (\pm)-**11** with fluorinated alkene.

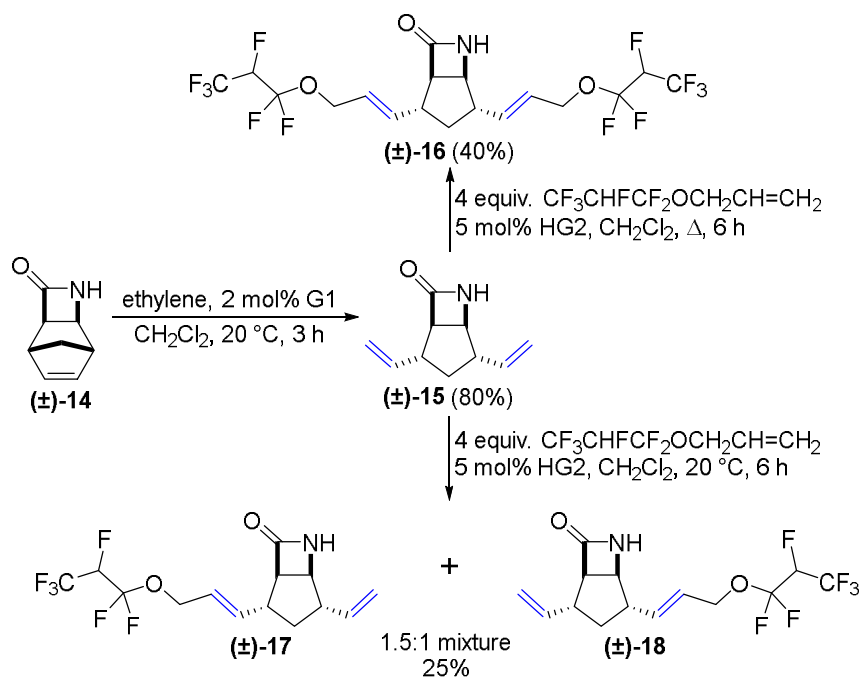
Two factors can contribute to the outcome of this reaction. First of all, chelation of the metallacycle intermediate with carbonyl oxygens is known to hinder further reactions by stabilizing the metallacycle [29-31]. During metathesis of the vinyl group attached directly to the lactam ring of (\pm)-**11**, formation of a 6-membered chelate ring with the oxo group of Boc is possible (Scheme 5, (\pm)-**T2** (\pm)-**T3**). As a result, reactivity of the C=C bond decreases. Another plausible factor is steric

hindrance of this vinyl group attached to the highly substituted azetidinone ring, which slows down its reaction with the Ru-alkylidene catalyst (Scheme 5).

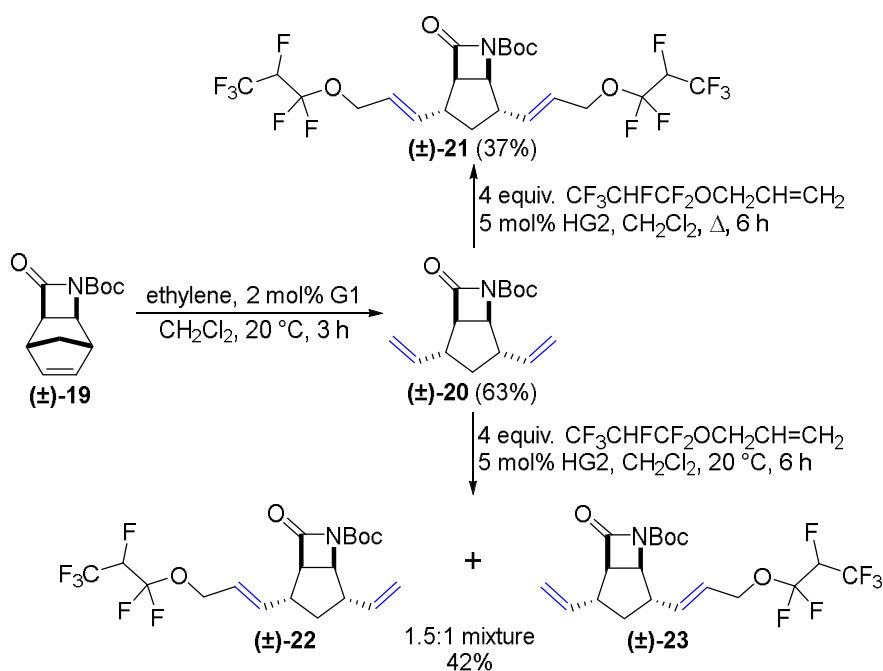


Scheme 5. Formation of mono-metathesized product (±)-13 and decoupled product (±)-12.

In continuation, lactam (±)-14 and its *N*-Boc-protected derivative (±)-19 were subjected to ROM according to our previous works [26, 27]. Cross metathesis of allyl 1,1,2,3,3,3-hexafluoropropyl ether with compounds (±)-15 and (±)-20 provided temperature-dependent results (Scheme 6 and Scheme 7). Under reflux in CH_2Cl_2 both (±)-15 and (±)-20 gave the corresponding dicoupled products (±)-16 and (±)-21. At room temperature, in turn, both (±)-15 and (±)-20 were transformed into 1.5:1 mixtures of two monocoupled products. This outcome is similar to our previous results on the chemoselectivity of CM reactions [26, 27]. Unfortunately, separation of the product mixtures in both cases was unsuccessful.



Scheme 6. Cross-metathesis of lactam $(\pm)\text{-15}$ with fluorinated alkene.



Scheme 7. Cross-metathesis of lactam $(\pm)\text{-20}$ with fluorinated alkene.

Conclusions

In the current work access to novel β -lactams, possessing fluorine atoms in their substituents, formed under cross-metathesis (CM) reaction conditions has been described. The synthetic concept involved the transformation of various diolefinated β -lactams across CM by using fluorine-containing alkene building elements. CM reaction of the dialkenylated β -lactams,

depending on their structure, furnished interesting di- or monocoupled functionalized derivatives. Stereochemical factors through chelate ring formation provide plausibly explanation for the outcome of the CM reactions. Further investigations in view of the synthesis through CM of novel fluorine-functionalized β -lactams are currently being studied in our laboratories.

Experimental

General information

Chemicals were purchased from Sigma-Aldrich. Solvents were used as received from the suppliers. Melting points were determined with a Kofler apparatus. Elemental analyses were performed with a Perkin–Elmer CHNS-2400 Ser II elemental analyser. Silica gel 60 F254 was purchased from Merck. NMR spectra were acquired at room temperature on a Bruker Avance 400 spectrometer (^1H frequency 400.13 MHz; ^{19}F frequency 376.50 MHz, ^{13}C frequency 100.76 MHz) or Bruker Avance Neo spectrometer (^1H frequency 500.20 MHz; ^{19}F frequency 470.66 MHz, ^{13}C frequency 125.78 MHz) in CDCl_3 or D_6 -DMSO solution, using the deuterium signal of the solvent to lock the field. The ^1H and ^{13}C chemical shifts are given relative to TMS and those of ^{19}F to CFCl_3 (0.00 ppm).

General Procedure for Ring-Opening Metathesis

β -Lactam (500 mg) was dissolved in anhydrous CH_2Cl_2 (100 ml) and Grubbs 1st generation catalyst (2 mol%) was added. The mixture was stirred at 20 °C under ethylene for 3 hours monitored by TLC. When the reaction was complete, the solution of NaHCO_3 (0.3 g) in H_2O (25 ml) and EtOH (5 ml) was added, and the mixture was stirred at 20 °C for another 2 hours with the aim of deactivating the catalyst. Then H_2O (30 ml) was added and the mixture was extracted with CH_2Cl_2 (3 \times 40 ml). The organic layer was dried (Na_2SO_4) and concentrated under reduced pressure. The crude material was purified by column chromatography on silica gel (*n*-hexane/EtOAc or *n*-hexane/acetone).

General Procedure for Cross Metathesis

β -Lactam (150 mg) was dissolved in anhydrous CH_2Cl_2 (30 ml) and then Hoveyda-Grubbs 2nd generation catalyst (5 mol%) and 3-(1,1,2,3,3,3-hexafluoropropoxy)prop-1-ene (4 equiv.) were added. The mixture was stirred at 40 °C under argon atmosphere for 6 hours monitored by TLC. When the reaction was complete, the solution of NaHCO_3 (0.3 g) in H_2O (25 ml) and EtOH (5 ml) was added, and the mixture was stirred at 20 °C another 2 hours (catalyst deactivation). After the

addition of H₂O (20 ml) the mixture was extracted with CH₂Cl₂ (3×30 ml). The organic layer was dried (Na₂SO₄) and concentrated under reduced pressure. The crude material was purified by column chromatography on silica gel (*n*-hexane/EtOAc or *n*-hexane/acetone).

General Procedure for the Preparation of N-Boc-Protected Lactams

β-Lactam (200 mg) was dissolved in THF (40 ml) then triethylamine (3 equiv.), di-*tert*-butyl dicarbonate (1.5 equiv.), and 4-dimethylaminopyridine (0.3 equiv.) were added at 0 °C. The mixture was stirred for 1 hour, then it was warmed to room temperature with stirring continued for 24 hours. When TLC indicated complete reaction, H₂O (30 ml) was added, and the mixture was extracted with EtOAc (3×20 ml). The organic layer was dried (Na₂SO₄) and concentrated under reduced pressure. The crude material was purified by column chromatography on silica gel (*n*-hexane/EtOAc 10:1).

For characterization and NMR data of the new compounds see Supporting Information.

Acknowledgments

We are grateful to the Hungarian Research Foundation (NKFIH No K 119282) for financial support. The financial support of the GINOP-2.3.2-15-2016-00038 project is also acknowledged. This research was supported by the EU-funded Hungarian grant EFOP-3.6.1-16-2016-00008. Ministry of Human Capacities, Hungary grant 20391-3/2018/FEKUSTRAT is also acknowledged.

References.

1. Kong, K.-F.; Schneper, L.; Mathee, K. *APMIS* **2010**, 118, 1-36.
2. Papp-Wallace, K. M.; Endimiani, A.; Taracila, M. A.; Bonomo, R. A. *Antimicrob. Agents Chemother.* **2011**, 55, 4943-4960.
3. Galletti, P.; Giacomini, D. *Curr. Med. Chem.* **2011**, 18, 4265-4283.
4. Veinberg, G.; Potorocina, I.; Vorona, M. *Curr. Med. Chem.* **2013**, 21, 393-416.
5. Kamath, A.; Ojima, I.; *Tetrahedron* **2012**, 68, 10640-10664.
6. Piens, N.; De Kimpe, N.; D'hooghe, M. *Prog. Heterocycl. Chem.* **2016**, 28, 27-55.
7. Dražić, T.; Roje, M. *Chem. Heterocycl. Compd.* **2017**, 53, 953-962.
8. a) Kiss, L.; Fülöp, F. *Chem. Rec.* **2018**, 18, 266-281; b) Kiss, L.; Mándity, I. M.; Fülöp, F. *Amino Acids* **2017**, 49, 1441-1455.
9. Zhou, Y.; Wang, Y.; Gu, Z.; Wang, S.; Zhu, W.; Acena, J. L.; Soloshonok, V. A.; Izawa, K.; Liu, H. *Chem. Rev.* **2016**, 116, 442-518.

10. Wang, J.; Sánchez-Roselló, M.; Aceña, J. L.; del Pozo, C.; Sorochinsky, A. E.; Fustero, S.; Soloshonok, V. A.; Liu, H. *Chem. Rev.* **2014**, 114, 2432-2506.
11. O'Hagan D. J. *Fluorine Chem.* **2010**, 131, 1071-1081.
12. a) Kiss, L.; Forró, E.; Fustero, S.; Fülöp, F. *Eur. J. Org. Chem.* **2011**, 4993-5001; b) Kiss, L.; Remete, A. M. *Eur. J. Org. Chem.* **2019**, 5574–5602.
13. Nonn, M.; Kiss, L.; Haukka, M.; Fustero, S.; Fülöp, F. *A Org. Lett.* **2015**, 17, 1074-1077 (and references therein).
14. Tarui, A.; Kondo, S.; Sato, K.; Omote, M.; Minami, H.; Miwa, Y.; Ando, A. *Tetrahedron* **2013**, 69, 1559-1565.
15. Sundell, R.; Siirola, E.; Kanerva, L. T. *Eur. J. Org. Chem.* **2014**, 6753-6760.
16. Trulli, L.; Raglione, V.; Fioravanti, S. *Eur. J. Org. Chem.* **2018**, 3743-3749.
17. Thi, H. D.; Thuy, G. L. N.; Catak, S.; Van Speybroeck, V.; Nguyen, T. V.; D'hooghe, M. *Synthesis* **2018**, 50, 1439-1456.
18. Thi, H. D.; Goossens, H.; Hertsen, D.; Otte, V.; Nguyen, T. V.; Van Speybroeck, V.; D'hooghe, M. *Chem. Asian J.* **2018**, 13, 421-431.
19. El Dine, A. N.; Grée, D.; Roisnel, T.; Caytan, E.; Hachem, A.; Grée, R. *Eur. J. Org. Chem.* **2016**, 556-561.
20. Kuznetsova, L.; Ungureanu, I. M.; Pepe, A.; Zanardi, I.; Wu, X.; Ojima, I. *J. Fluorine Chem.* **2004**, 125, 487-500.
21. Joyeau, R.; Felk, A.; Guillaume, S.; Wakselman, M.; Vergely, I.; Doucet, C.; Boggetto, N.; Reboud-Ravaux, M. *J. Pharm. Pharmacol.* **1996**, 48, 1218-1230.
22. Higman, C. S.; Lummiss, J. A. M.; Fogg, D. E. *Angew. Chem. Int. Ed.* **2016**, 55, 3552-3565.
23. Monsaert, S.; Vila, A. L.; Drozdak, R.; Van Der Voort, P.; Verpoort, F. *Chem. Soc. Rev.* **2009**, 38, 3360-3372.
24. Prunet, J.; *Eur. J. Org. Chem.* **2011**, 3634-3647.
25. Smith, B. J.; Sulikowski, G. A. *Angew. Chem. Int. Ed.* **2010**, 49, 1599-1602.
26. Kardos, M.; Kiss, L.; Fülöp, F. *Asian. J. Org. Chem.* **2015**, 4, 1155-1159.
27. Kardos, M.; Kiss, L.; Haukka, M.; Fustero, S.; Fülöp, F. *Eur. J. Org. Chem.* **2017**, 1894-1901.
28. Fülöp, F.; Forró, E.; Tóth, G. K. *Org. Lett.* **2004**, 6, 4239-4241.
29. BouzBouz, S.; Cossy, J. *Org. Lett.* **2001**, 3, 1451-1454.
30. Boufroura, H.; Mauduit, M.; Drège, E.; Joseph, D. *J. Org. Chem.* **2013**, 78, 2346-2354.
31. Nonn, M.; Benke, Z.; Fustero, S.; Fülöp, F.; Kiss, L. *Eur. J. Org. Chem.* **2019**, 5285–5293.

Supporting Information

STEREOCONTROLLED SYNTHESIS OF FLUORINE-CONTAINING FUNCTIONALIZED B-LACTAM DERIVATIVES THROUGH CROSS- METATHESIS REACTION

Attila M. Remete,^{1,2} Zsanett Benke,^{1,2} Loránd Kiss^{1,2*}

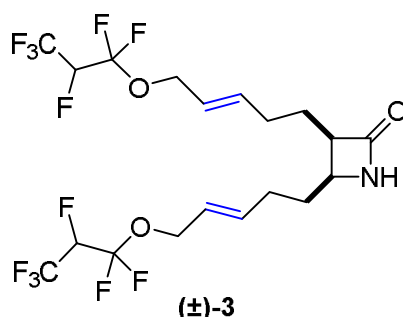
¹*Institute of Pharmaceutical Chemistry, University of Szeged, H-6720 Szeged, Eötvös u. 6, Hungary*

²*University of Szeged, Interdisciplinary Excellence Centre, Institute of Pharmaceutical Chemistry, H-6720 Szeged, Eötvös u. 6, Hungary*

E-mail: kiss.lorand@pharm.u-szeged.hu

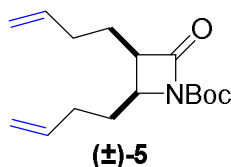
Characterization of the synthesized new substances

(3*R**,4*S**)-3,4-bis((*E*)-5-(1,1,2,3,3,3-Hexafluoropropoxy)pent-3-en-1-yl)azetidin-2-one



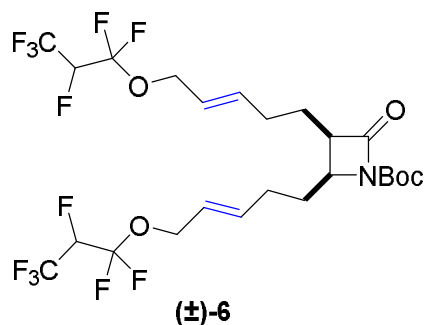
Light brown oil, yield: 26%, $R_f = 0.59$ (n-hexane/EtOAc 1:3); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ (ppm): 1.54-1.73 (m, 3H, CH_2), 1.77-1.85 (m, 1H, CH_2), 2.10-2.27 (m, 3H, CH_2), 2.30-2.42 (m, 1H, CH_2) 3.16-3.25 (m, 1H, H-3), 3.63-3.71 (m, 1H, H-4), 4.42-4.48 (m, 4H, OCH_2), 4.67-4.88 (m, 2H, CFH), 5.56-5.67 (m, 2H, $=\text{CH}$), 5.75-5.86 (m, 2H, $=\text{CH}$), 5.90 (brs, 1H, NH); $^{19}\text{F NMR}$ (471 MHz, CDCl_3) δ (ppm): -75.14, -79.82, -82.33, -211.39; $^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ (ppm): 24.1, 29.6, 30.1, 30.2, 51.7, 52.3, 65.0 (t, $^3\text{J} = 6.2$ Hz, OCH_2), 65.2 (t, $^3\text{J} = 6.1$ Hz, OCH_2), 84.8 (doublet sextet, $^1\text{J} = 201.1$ Hz, $^2\text{J} = 35.9$ Hz, CFH), 84.8 (doublet sextet, $^1\text{J} = 200.6$ Hz, $^2\text{J} = 35.9$ Hz, CFH), 118.5 (td, 2C, $^1\text{J} = 268.1$ Hz, $^2\text{J} = 24.7$ Hz, CF_2), 120.0 (qd, 2C, $^1\text{J} = 282.4$ Hz, $^2\text{J} = 24.9$ Hz, CF_3), 124.2, 124.4, 135.1, 135.5, 171.3.

(2*S**,3*R**)-tert-Butyl 2,3-di(but-3-en-1-yl)-4-oxoazetidine-1-carboxylate



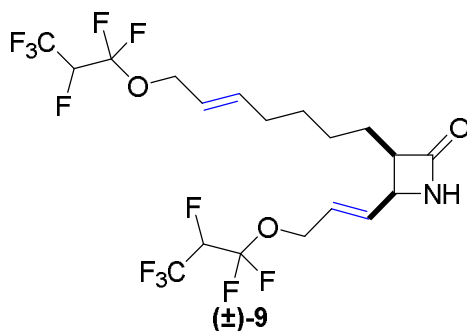
Colorless oil, yield: 76%, $R_f = 0.59$ (n-hexane/EtOAc 4:1); $^1\text{H NMR}$ (500 MHz, CDCl_3) δ (ppm): 1.52 (s, 9H, CH_3), 1.62-1.76 (m, 2H, CH_2), 1.79-1.90 (m, 1H, CH_2), 2.00-2.10 (m, 1H, CH_2) 2.13-2.27 (m, 3H, CH_2), 2.32-2.43 (m, 1H, CH_2) 3.22-3.29 (m, 2H, H-3), 3.97-4.04 (m, 1H, H-2), 4.99-5.11 (m, 4H, $=\text{CH}_2$), 5.74-5.88 (m, 2H, $=\text{CH}$); $^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ (ppm) = 23.8, 28.1, 28.1, 30.7, 31.6, 51.0, 54.5, 83.1, 115.4, 115.9, 137.2, 137.2, 148.4, 168.3.

(2*S**,3*R**)-tert-Butyl 2,3-bis((*E*)-5-(1,1,2,3,3,3-hexafluoropropoxy)pent-3-en-1-yl)-4-oxoazetidine-1-carboxylate



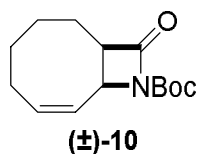
Light brown oil, yield: 34%, $R_f = 0.41$ (n-hexane/EtOAc 4:1); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ (ppm): 1.52 (s, 9H, CH_3), 1.59-1.76 (m, 2H, CH_2), 1.78-1.91 (m, 1H, CH_2), 1.97-2.11 (m, 1H, CH_2), 2.14-2.32 (m, 3H, CH_2), 2.35-2.49 (m, 1H, CH_2), 3.17-3.28 (m, 1H, H-3), 3.94-4.05 (m, 1H, H-2), 4.40-4.50 (m, 4H, OCH_2), 4.67-4.88 (m, 2H, CFH), 5.57-5.69 (m, 2H, =CH), 5.73-5.88 (m, 2H, =CH); $^{19}\text{F NMR}$ (471 MHz, CDCl_3) δ (ppm) = -75.20, -79.88, -82.42, -211.46; $^{19}\text{F NMR}$ (471 MHz, CDCl_3) δ (ppm): -79.9, -82.2, -82.3, -82.4, -211.4; $^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ (ppm): 23.8, 28.1, 28.2, 29.2, 30.0, 50.8, 54.3, 65.1 (t, $^3\text{J} = 6.0$ Hz, OCH_2), 65.1 (t, $^3\text{J} = 6.0$ Hz, OCH_2), 83.4, 84.8 (doublet sextet, 2C, $^1\text{J} = 200.8$ Hz, $^2\text{J} = 36.1$ Hz, CFH), 118.5 (td, 2C, $^1\text{J} = 268.4$ Hz, $^2\text{J} = 22.2$ Hz, CF_2), 120.0 (qd, 2C, $^1\text{J} = 282.0$ Hz, $^2\text{J} = 26.4$ Hz, CF_3), 124.2, 124.6, 134.9, 135.0, 135.1, 148.4, 167.9.

(3*R,4*S**)-3-((*E*)-7-(1,1,2,3,3,3-Hexafluoropropoxy)hept-5-en-1-yl)-4-((*E*)-3-(1,1,2,3,3,3-hexafluoropropoxy)prop-1-en-1-yl)azetidin-2-one**



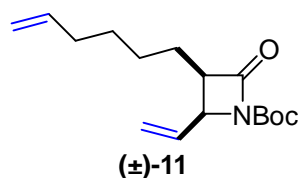
Light brown oil, yield: 35%, $R_f = 0.45$ (n-hexane/acetone 2:1); $^1\text{H NMR}$ (500 MHz, CDCl_3) δ (ppm): 1.30-1.53 (m, 4H, CH_2), 1.55-1.75 (m, 2H, CH_2), 2.00-2.12 (m, 2H, CH_2), 3.26-3.36 (m, 1H, H-3), 4.23-4.30 (m, 1H, H-4), 4.40-4.46 (m, 2H, OCH_2), 4.50-4.58 (m, 2H, OCH_2), 4.68-4.92 (m, 2H, CFH), 5.49-5.59 (m, 1H, =CH), 5.72-5.83 (m, 1H, =CH), 5.83-5.90 (m, 2H, =CH), 6.01 (s, 1H, NH); $^{19}\text{F NMR}$ (471 MHz, CDCl_3) δ (ppm): -75.18, -79.78, -79.87, -82.29, -82.40, -211.45; $^{13}\text{C NMR}$ (126 MHz, DMSO) δ (ppm): 25.2, 26.9, 28.6, 31.7, 51.2, 54.7, 64.8-65.0 (m, OCH_2), 65.8 (t, $^3\text{J} = 5.8$ Hz, OCH_2), 84.3 (doublet sextet, 2C, $^1\text{J} = 193.8$ Hz, $^2\text{J} = 35.0$ Hz, CFH), 119.1 (td, 2C, $^1\text{J} = 267.4$ Hz, $^2\text{J} = 22.3$ Hz, CF_2), 120.8 (qd, 1C, $^1\text{J} = 281.8$ Hz, $^2\text{J} = 25.5$ Hz, CF_3), 120.8 (qd, 1C, $^1\text{J} = 281.3$ Hz, $^2\text{J} = 25.2$ Hz, CF_3), 123.7, 126.7, 133.2, 137.1, 170.5.

(1*R,8*S**,*Z*)-*tert*-Butyl 10-oxo-9-azabicyclo[6.2.0]dec-6-ene-9-carboxylate**



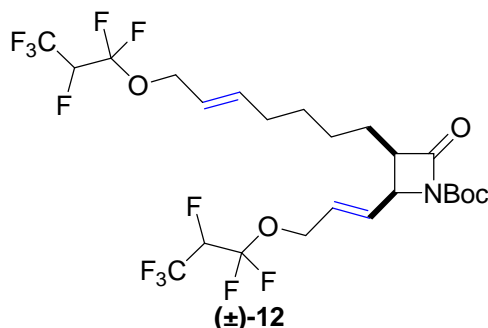
White solid, m.p. 60-74 °C, yield: 96%, $R_f = 0.49$ (n-hexane/EtOAc 4:1); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ (ppm): 1.36-1.49 (m, 2H, CH_2), 1.53 (s, 9H, CH_3), 1.62-1.83 (m, 2H, CH_2), 1.84-1.99 (m, 2H, CH_2), 2.01-2.15 (m, 2H, CH_2), 3.26-3.35 (m, 1H, H-1), 4.79-4.86 (m, 1H, H-8), 5.54-5.63 (m, 1H, =CH), 5.76-5.86 (m, 1H, =CH); $^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ (ppm): 22.0, 25.6, 28.1, 29.1, 31.2, 54.5, 56.7, 83.2, 123.5, 134.1, 147.8, 167.8.

(3*R,4*S**)-*tert*-Butyl 3-(hex-5-en-1-yl)-2-oxo-4-vinylazetidine-1-carboxylate**



Colorless oil, yield: 91%, $R_f = 0.51$ (n-hexane/EtOAc 4:1); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ (ppm): 1.30-1.44 (m, 4H, CH_2), 1.50 (s, 9H, CH_3), 1.52-1.60 (m, 1H, CH_2), 1.62-1.75 (m, 1H, CH_2), 1.97-2.10 (m, 2H, CH_2), 3.22-3.33 (m, 1H, H-3), 4.46 (t, 1H, $J = 7.13$ Hz, H-4), 4.89-5.04 (m, 2H, = CH_2), 5.33-5.43 (m, 2H, = CH_2), 5.71-5.88 (m, 2H, =CH); $^{13}\text{C NMR}$ (126 MHz, DMSO) δ (ppm) = 24.6, 26.8, 28.1, 28.4, 33.4, 53.0, 56.8, 82.4, 115.3, 119.9, 133.7, 139.0, 147.7, 168.2.

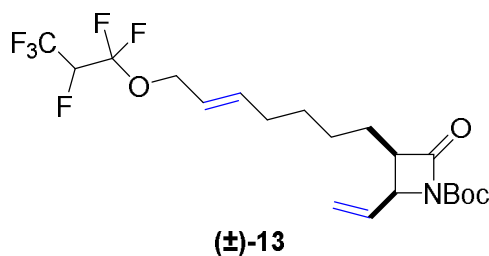
(2*S,3*R**)-*tert*-Butyl 3-((*E*)-7-(1,1,2,3,3,3-hexafluoropropoxy)hept-5-en-1-yl)-2-((*E*)-3-(1,1,2,3,3,3-hexafluoropropoxy)prop-1-en-1-yl)-4-oxoazetidine-1-carboxylate**



Yellow oil, yield: 16%, $R_f = 0.36$ (n-hexane/EtOAc 4:1); $^1\text{H NMR}$ (500 MHz, CDCl_3) δ (ppm): 1.28-1.44 (m, 5H, CH_2), 1.49 (s, 9H, CH_3), 1.60-1.72 (m, 1H, CH_2), 2.01-2.11 (m, 2H, CH_2), 3.25-3.33 (m, 1H, H-3), 4.43 (d, 2H, $J = 6.35$ Hz, OCH_2), 4.48-4.54 (m, 1H, H-2), 4.56 (d, 2H, $J = 4.27$ Hz, OCH_2), 4.70-4.88 (m, 2H, CFH), 5.50-5.58 (m, 1H, =CH), 5.72-5.84 (m, 2H, =CH), 5.85-5.94 (m, 1H, =CH); $^{19}\text{F NMR}$

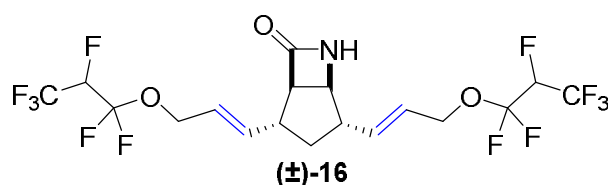
(471 MHz, CDCl₃) δ (ppm): -75.16, -79.75, -79.95, -82.14, -82.39, -211.44; ¹³C NMR (125 MHz, CDCl₃) δ (ppm): 24.8, 26.9, 28.0, 28.4, 31.7, 53.6, 55.4, 63.8-64.0 (m, OCH₂), 65.4 (t, ³J = 6.2 Hz, OCH₂), 83.4, 84.7 (doublet sextet, 2C, ¹J = 204.3 Hz, ²J = 37.7 Hz, CFH), 116.5-123.5 (m, 4C, CF₂ and CF₃), 123.4, 128.7, 128.8, 136.7, 147.8, 167.5.

(3R*,4S*)-tert-Butyl 3-((E)-7-(1,1,2,3,3,3-hexafluoropropoxy)hept-5-en-1-yl)-2-oxo-4-vinylazetidine-1-carboxylate



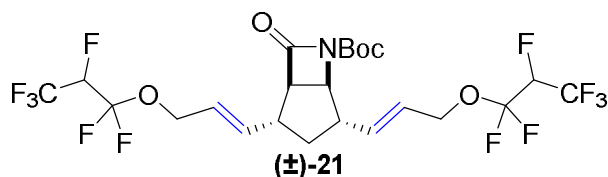
Yellow oil, yield: 25%, R_f = 0.45 (n-hexane/EtOAc 4:1); ¹H NMR (500 MHz, CDCl₃) δ (ppm): 1.32-1.45 (m, 4H, CH₂), 1.45-1.57 (m, 10H, Boc and CH₂), 1.57-1.73 (m, 1H, CH₂), 2.00-2.13 (m, 1H, CH₂), 3.22-3.32 (m, 1H, H-3), 4.39-4.50 (m, 3H, OCH₂ and H-4), 4.70-4.88 (m, 1H, CHF), 5.34-5.43 (m, 1H, =CH₂), 5.50-5.59 (m, 1H, =CH), 5.72-5.88 (m, 2H, =CH); ¹⁹F NMR (471 MHz, CDCl₃) δ (ppm) = -75.14, -79.77, -82.39, -211.40; ¹³C NMR (126 MHz, CDCl₃) δ (ppm): 24.8, 26.8, 28.0, 28.5, 31.8, 53.3, 56.8, 65.4 (t, 2H, J = 6.1 Hz, OCH₂), 83.2, 84.8 (doublet sextet, ¹J = 201.0 Hz, ²J = 36.7 Hz, CFH), 115.4-120.7 (m, 2C, CF₂ and CF₃), 120.3, 123.3, 132.2, 136.8, 147.9, 167.9.

(1R*,2R*,4S*,5S*)-2,4-bis((E)-3-(1,1,2,3,3,3-Hexafluoropropoxy)prop-1-en-1-yl)-6-azabicyclo[3.2.0]heptan-7-one



Light brown oil, yield: 40%, R_f = 0.48 (n-hexane/acetone 2:1); ¹H NMR (500 MHz, CDCl₃) δ (ppm): 1.89 (d, J = 13.89 Hz, 1H, H-3), 2.46-2.59 (m, 1H, H-3), 2.81 (t, J = 7.13 Hz, 1H, H-2), 3.10 (t, J = 7.00 Hz, 1H, H-4), 3.59 (s, 1H, H-1), 4.04 (d, J = 3.21 Hz, 1H, H-5), 4.38-4.47 (m, 4H, OCH₂), 4.68-4.87 (m, 2H, CFH), 5.53-5.65 (m, 2H, =CH), 5.66-5.77 (m, 2H, =CH), 5.81 (brs, 1H, NH); ¹⁹F NMR (471 MHz, CDCl₃) δ (ppm): -75.22, -79.99, -82.21, -211.45; ¹³C NMR (126 MHz, DMSO) δ (ppm): 37.1, 38.5, 44.3, 58.1, 60.6, 65.6 (t, ³J = 6.3 Hz, OCH₂), 65.6 (t, ³J = 6.2 Hz, OCH₂), 84.3 (doublet sextet, 2C, ¹J = 193.8 Hz, ²J = 35.2 Hz, CFH), 119.1 (td, 2C, ¹J = 266.5 Hz, ²J = 23.2 Hz, CF₂), 120.8 (qd, 2C, ¹J = 281.1 Hz, ²J = 25.1 Hz, CF₃), 122.9, 124.0, 138.2, 139.9, 168.9.

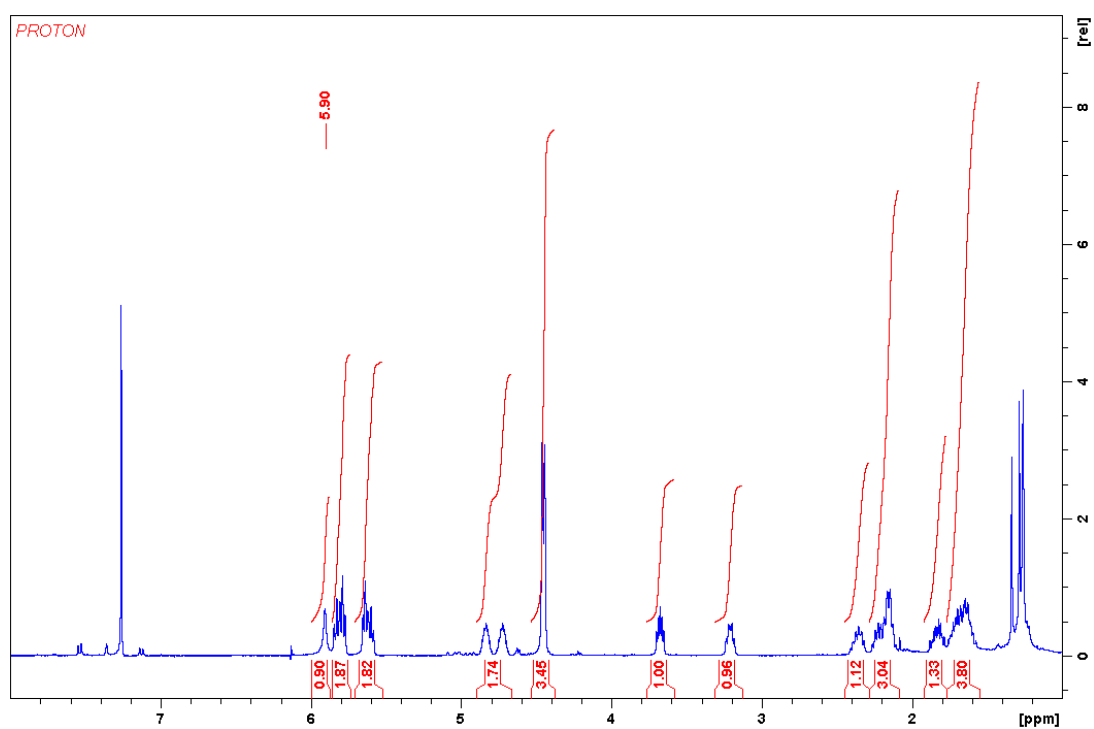
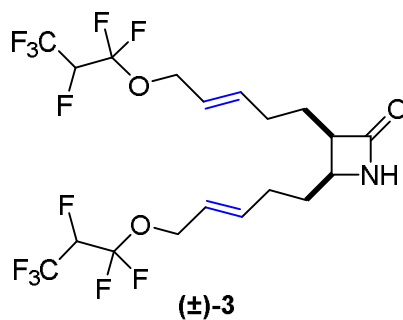
(1*R,2*R**,4*S**,5*S**)-tert-Butyl 2,4-bis((*E*)-3-(1,1,2,3,3,3-hexafluoropropoxy)prop-1-en-1-yl)-7-oxo-6-azabicyclo[3.2.0]heptane-6-carboxylate**

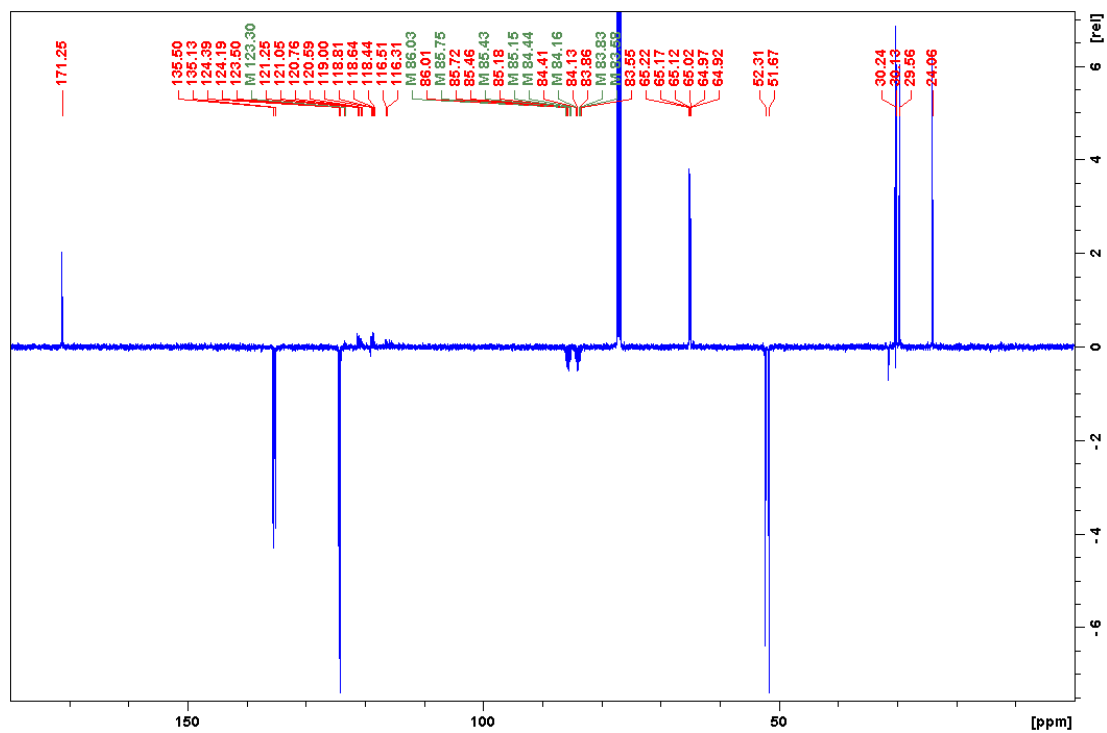


Yellow oil, yield: 37%, $R_f = 0.40$ (n-hexane/EtOAc 5:1); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ (ppm): 1.52 (s, 9H, CH_3), 1.84-1.93 (m, 1H, H-3), 2.33-2.48 (m, 1H, H-3), 3.05-3.17 (m, 2H, H-2, H-4), 3.48-3.54 (m, 1H, H-1) 4.26 (d, 1H, $J = 4.5$ Hz, H-5), 4.40-4.47 (m, 4H, OCH_2), 4.68-4.88 (m, 2H, CFH), 5.55-5.82 (m, 4H, =CH); $^{19}\text{F NMR}$ (471 MHz, CDCl_3) δ (ppm): -75.21, -80.03, -82.26, -211.45; $^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ (ppm): 28.0, 37.2, 40.4, 43.1, 59.3, 62.4, 64.6 (t, $^3J = 6.2$ Hz, OCH_2), 64.7 (t, $^3J = 6.2$ Hz, OCH_2), 83.7, 84.7 (doublet sextet, 2C, $^1J = 201.0$ Hz, $^2J = 36.2$ Hz, CFH), 118.5 (td, 2C, $^1J = 268.3$ Hz, $^2J = 24.2$ Hz, CF_2), 120.0 (qd, 2C, $^1J = 281.5$ Hz, $^2J = 25.7$ Hz, CF_3), 123.7, 124.7, 136.1, 137.6, 147.5, 166.0.

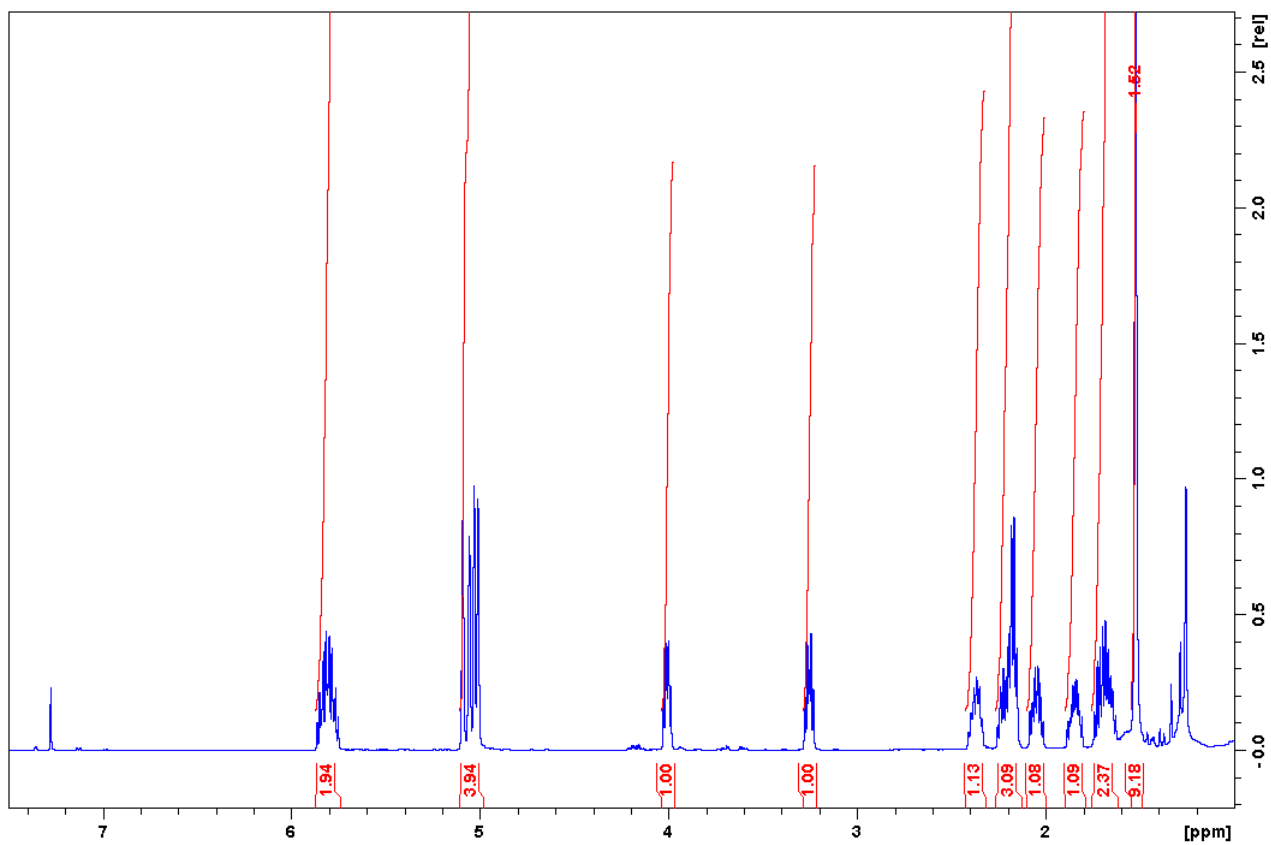
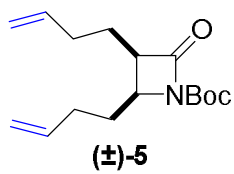
^1H and ^{13}C NMR spectra of the synthesized new substances

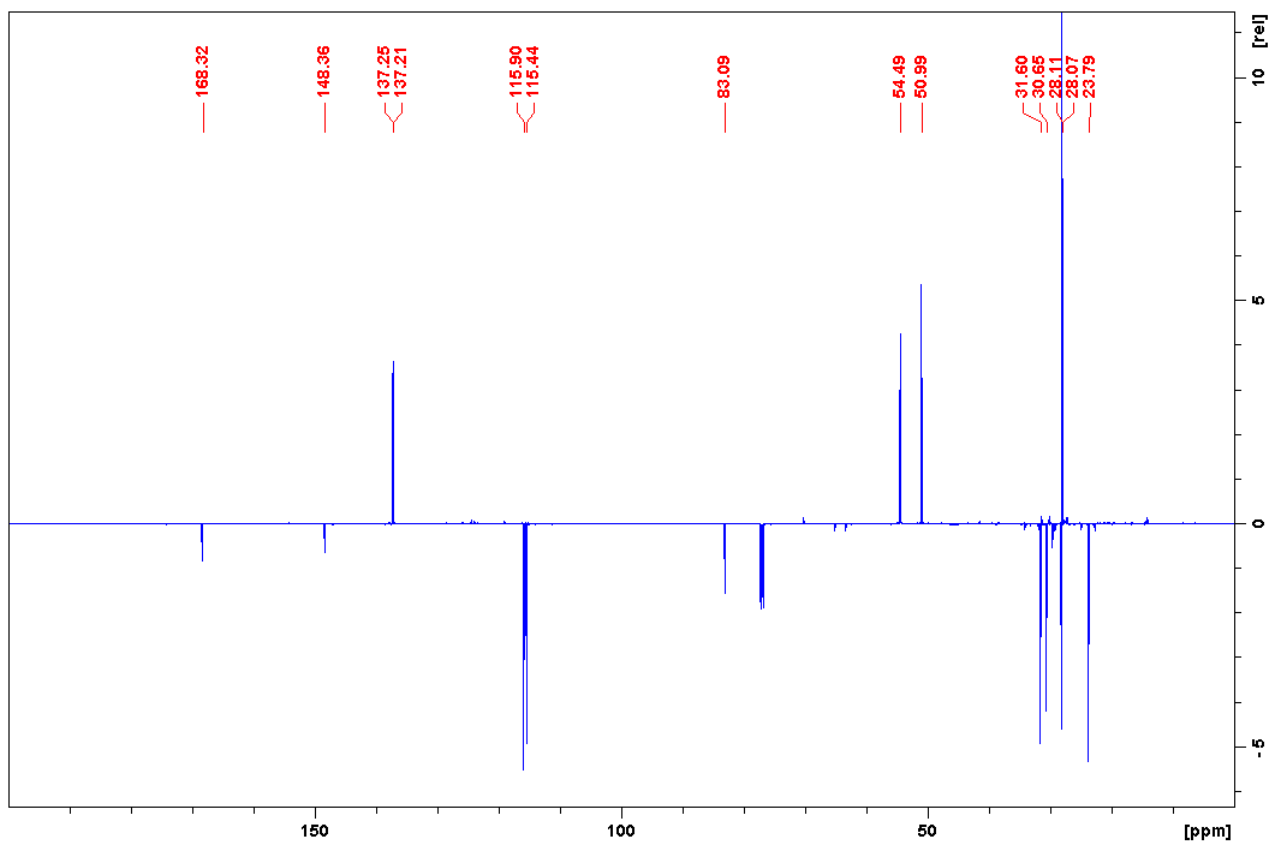
(3*R,4*S**)-3,4-bis((*E*)-5-(1,1,2,3,3,3-Hexafluoropropoxy)pent-3-en-1-yl)azetidin-2-one**



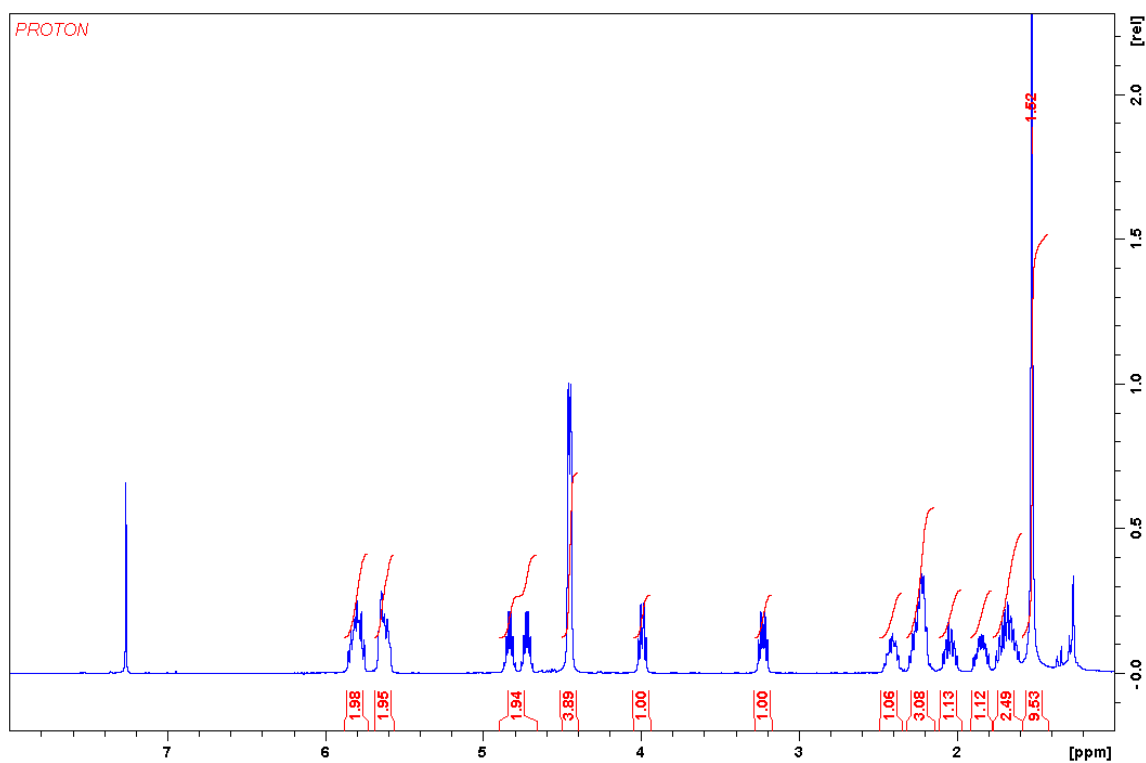
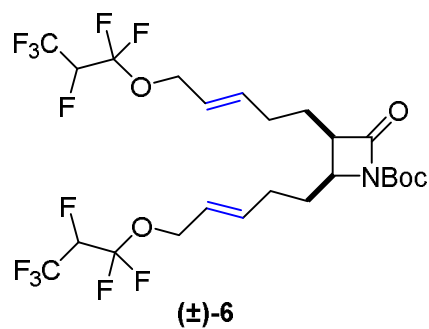


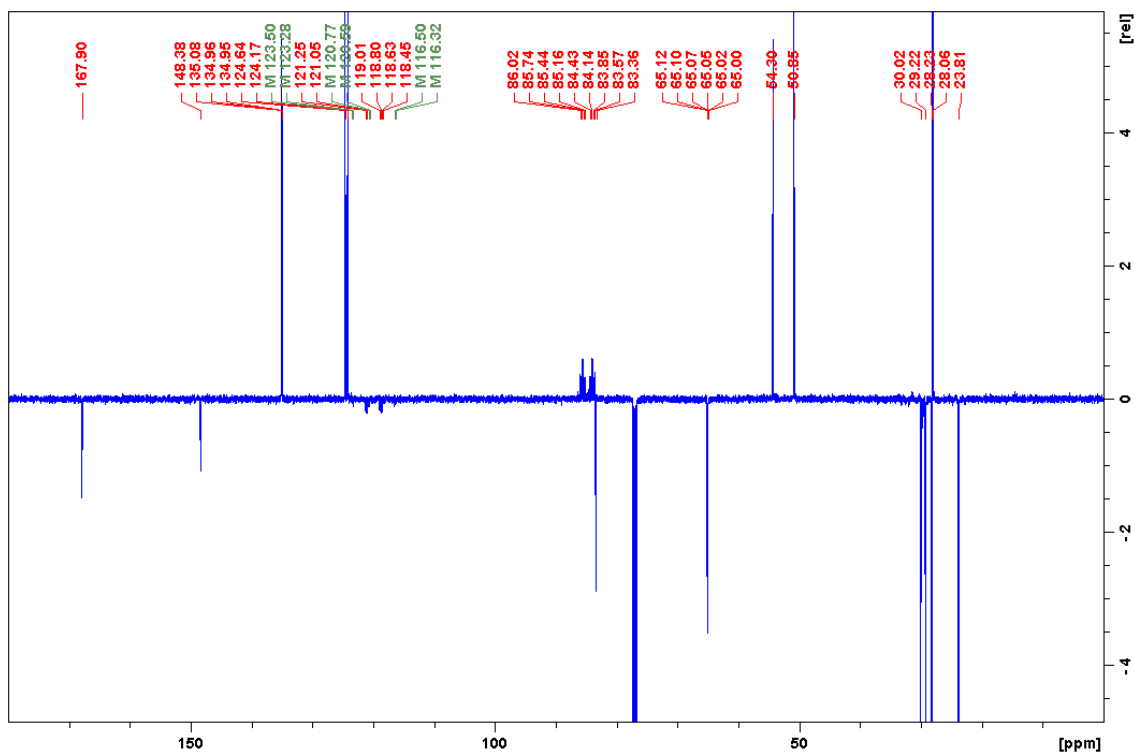
(2*S,3*R**)-tert-Butyl 2,3-di(but-3-en-1-yl)-4-oxoazetidine-1-carboxylate**



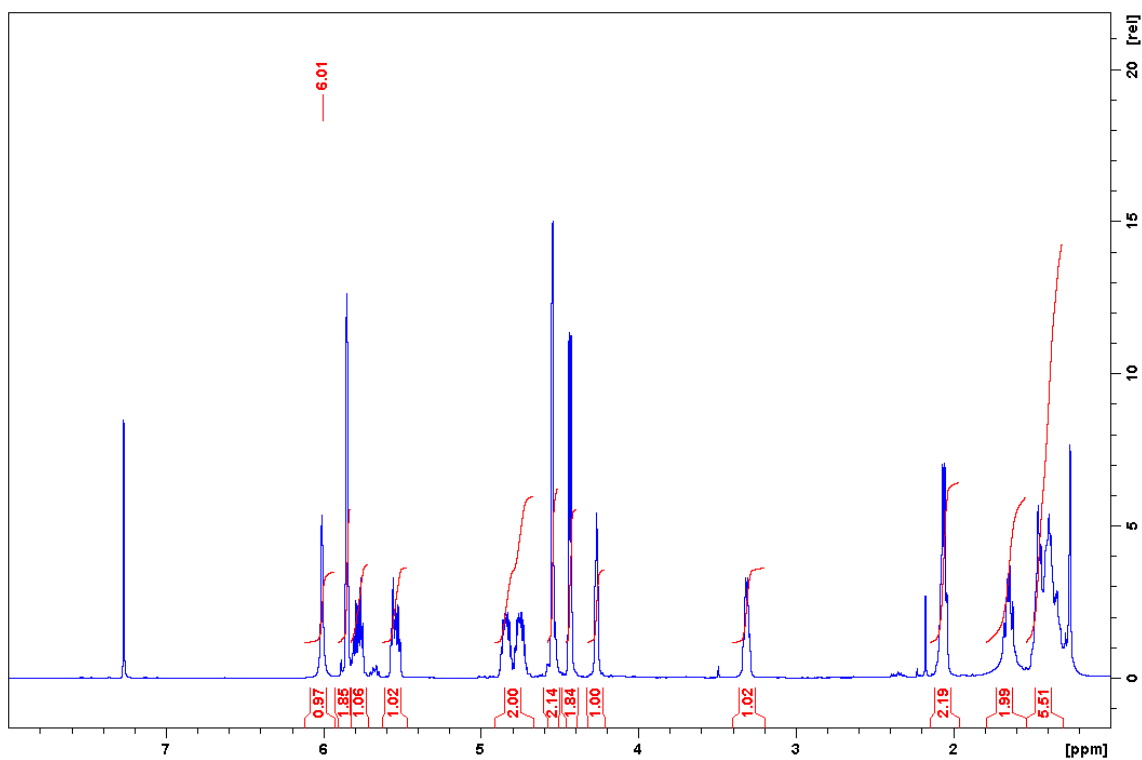
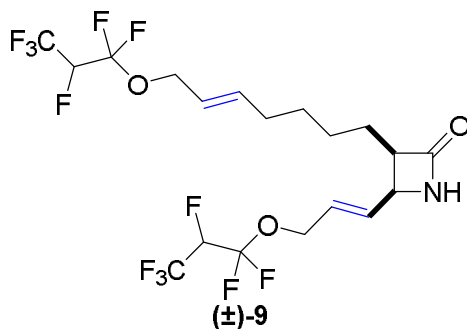


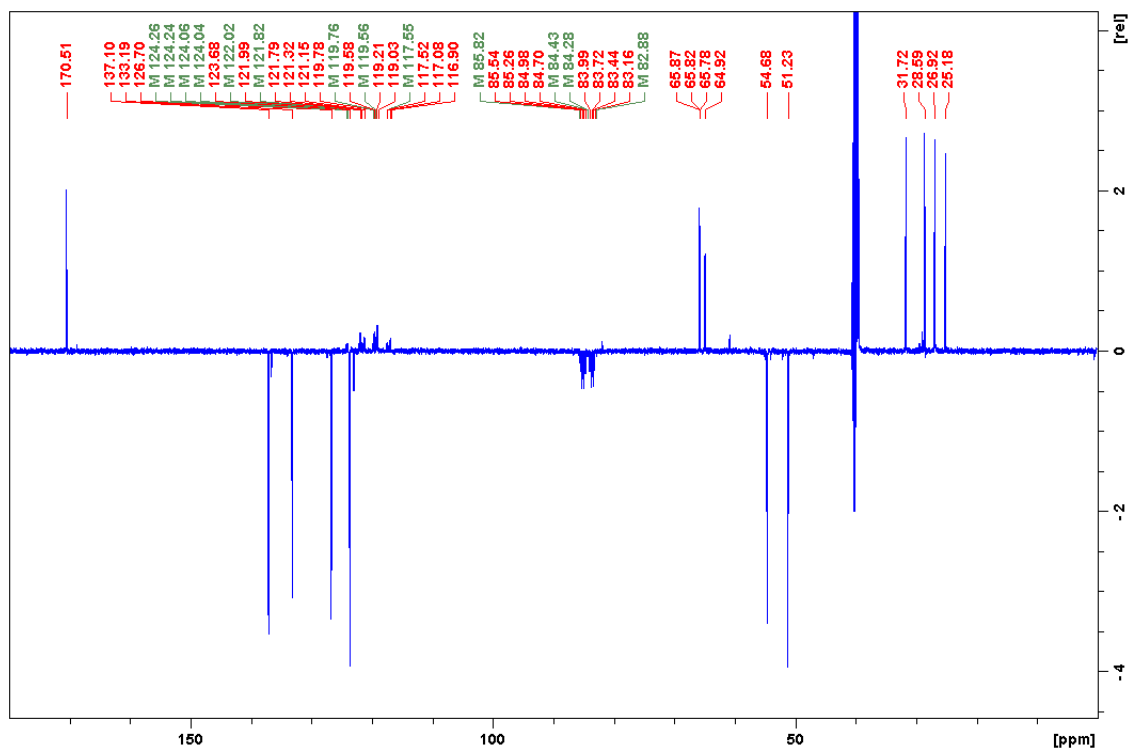
(2*S,3*R**)-*tert*-Butyl 2,3-bis((*E*)-5-(1,1,2,3,3,3-hexafluoropropoxy)pent-3-en-1-yl)-4-oxoazetidine-1-carboxylate**



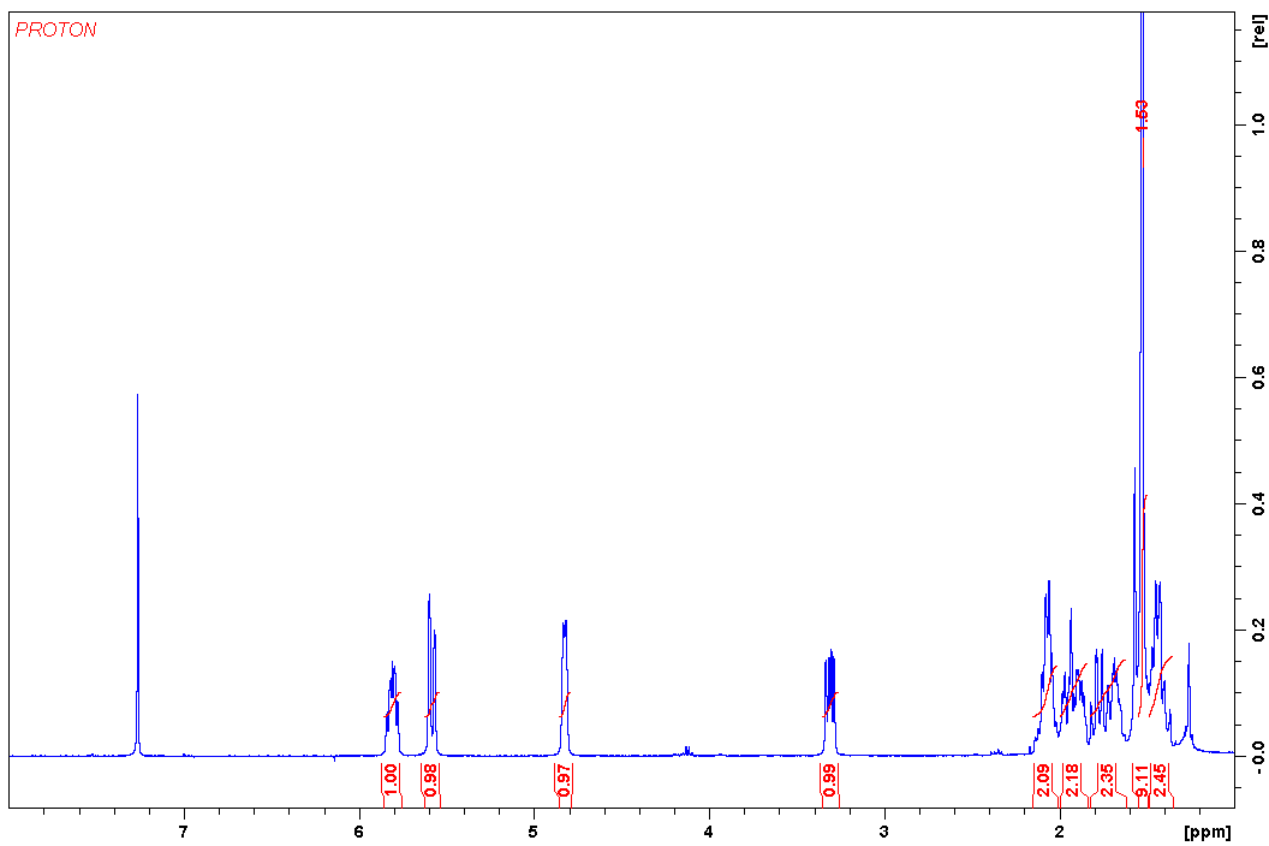
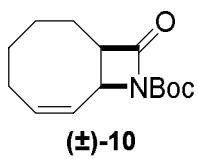


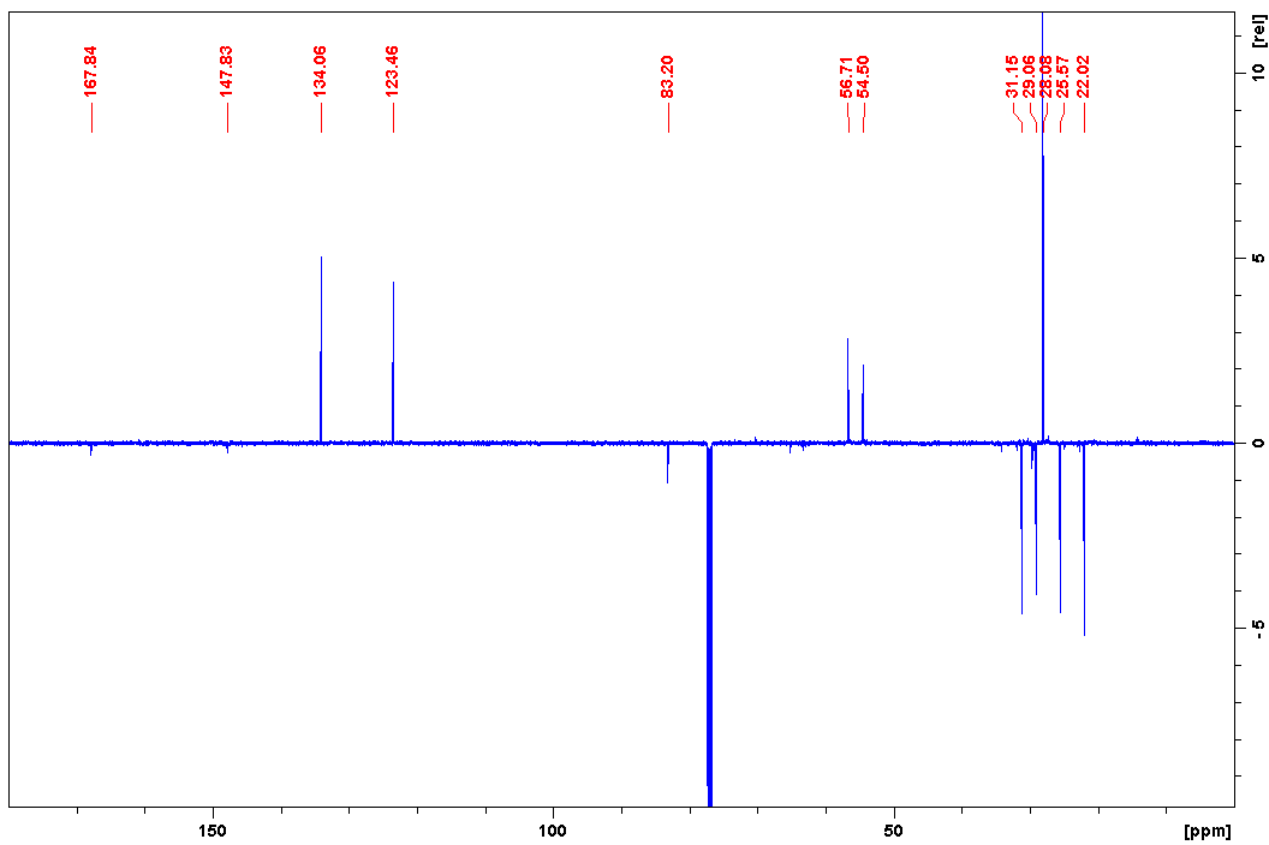
(3*R,4*S**)-3-((*E*)-7-(1,1,2,3,3,3-Hexafluoropropoxy)hept-5-en-1-yl)-4-((*E*)-3-(1,1,2,3,3,3-hexafluoropropoxy)prop-1-en-1-yl)azetidin-2-one**



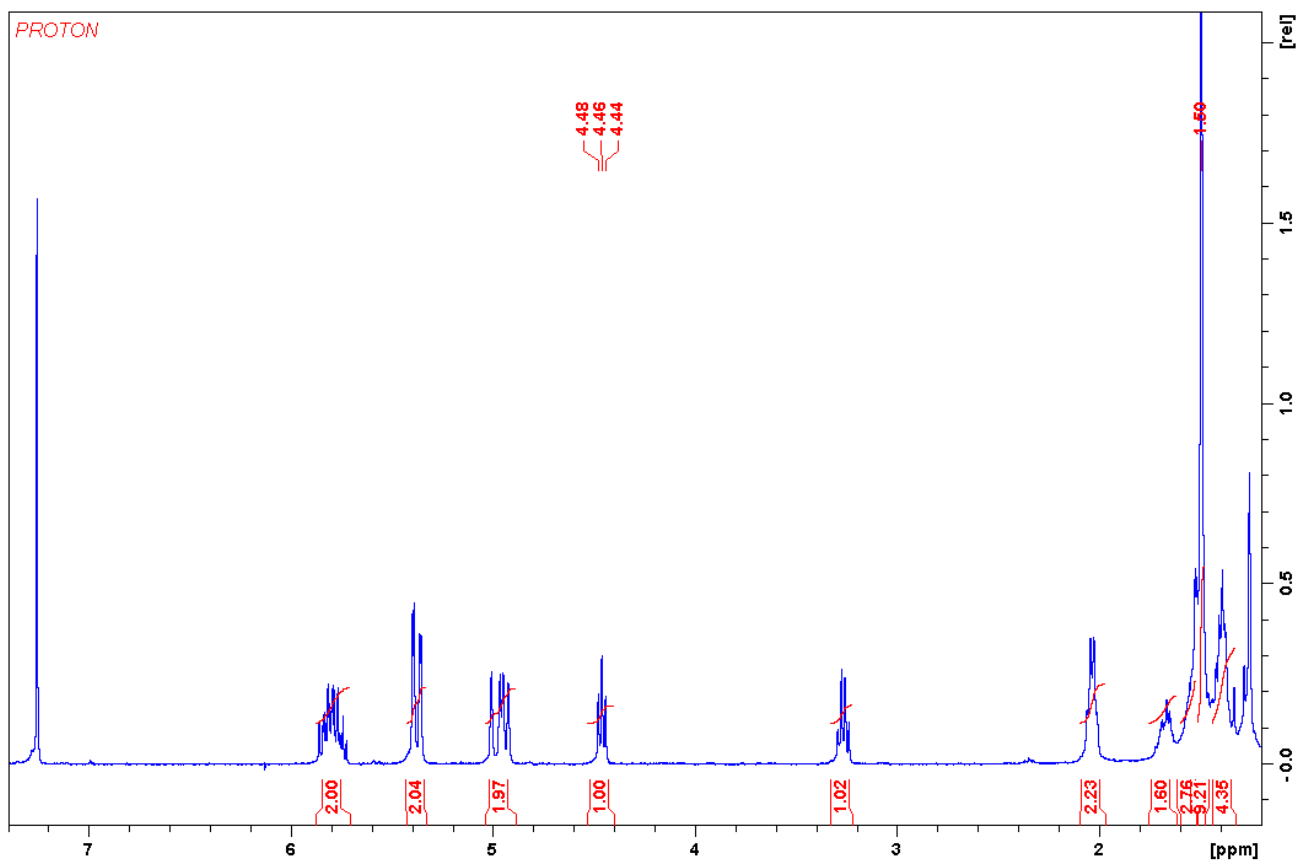
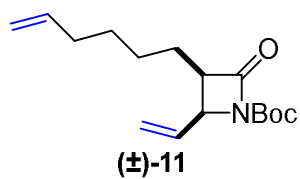


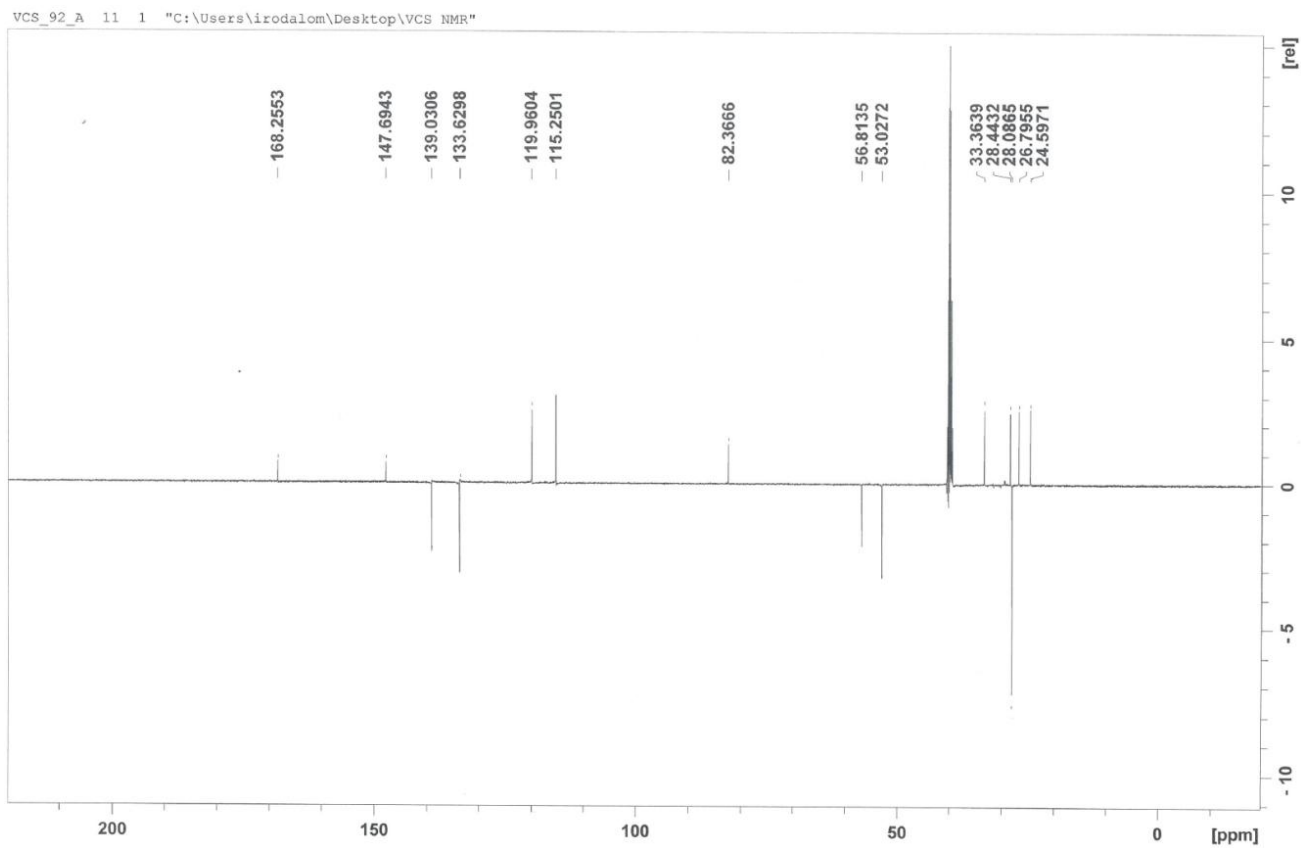
(1*R,8*S**,*Z*)-tert-Butyl 10-oxo-9-azabicyclo[6.2.0]dec-6-ene-9-carboxylate**



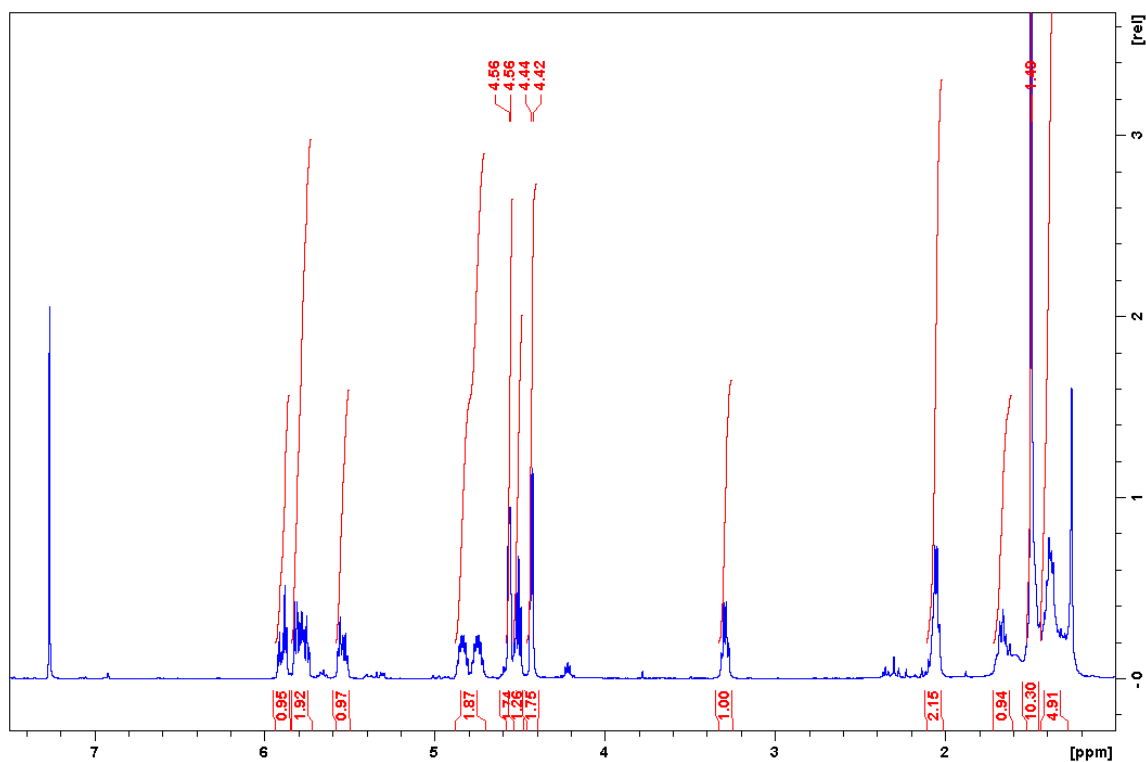
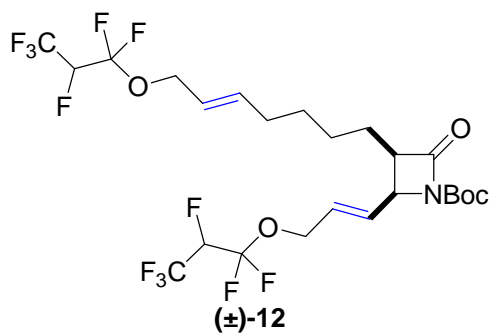


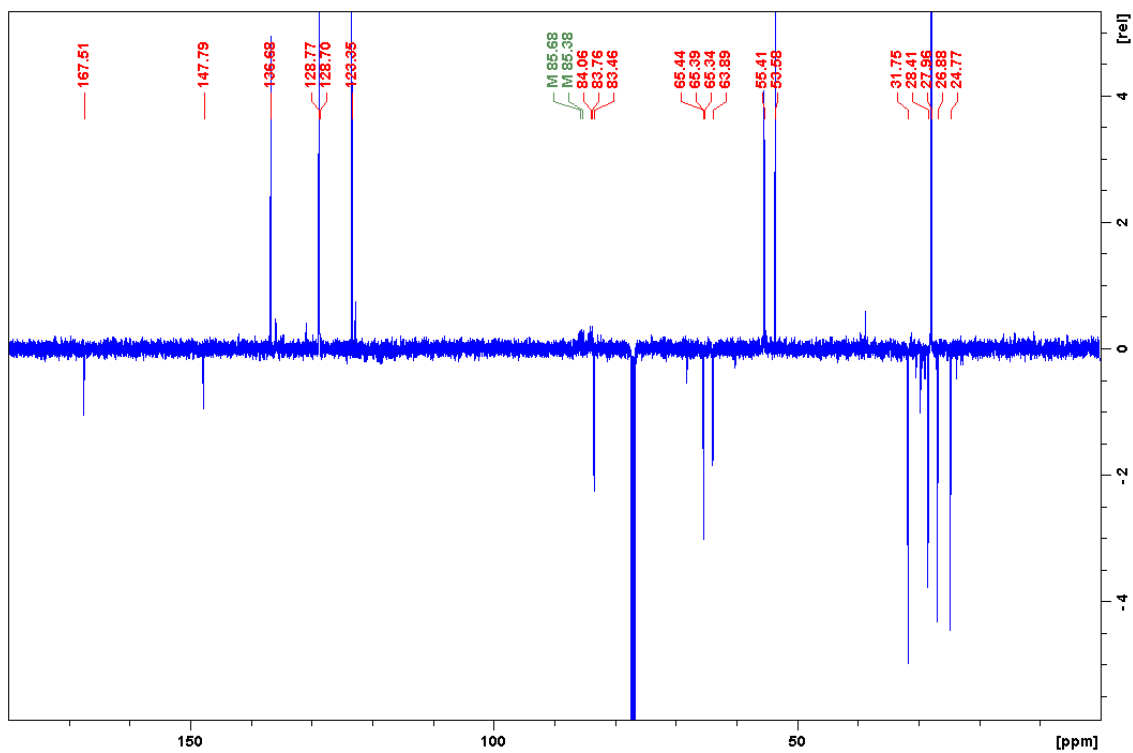
(3*R,4*S**)-*tert*-Butyl 3-(hex-5-en-1-yl)-2-oxo-4-vinylazetidine-1-carboxylate**



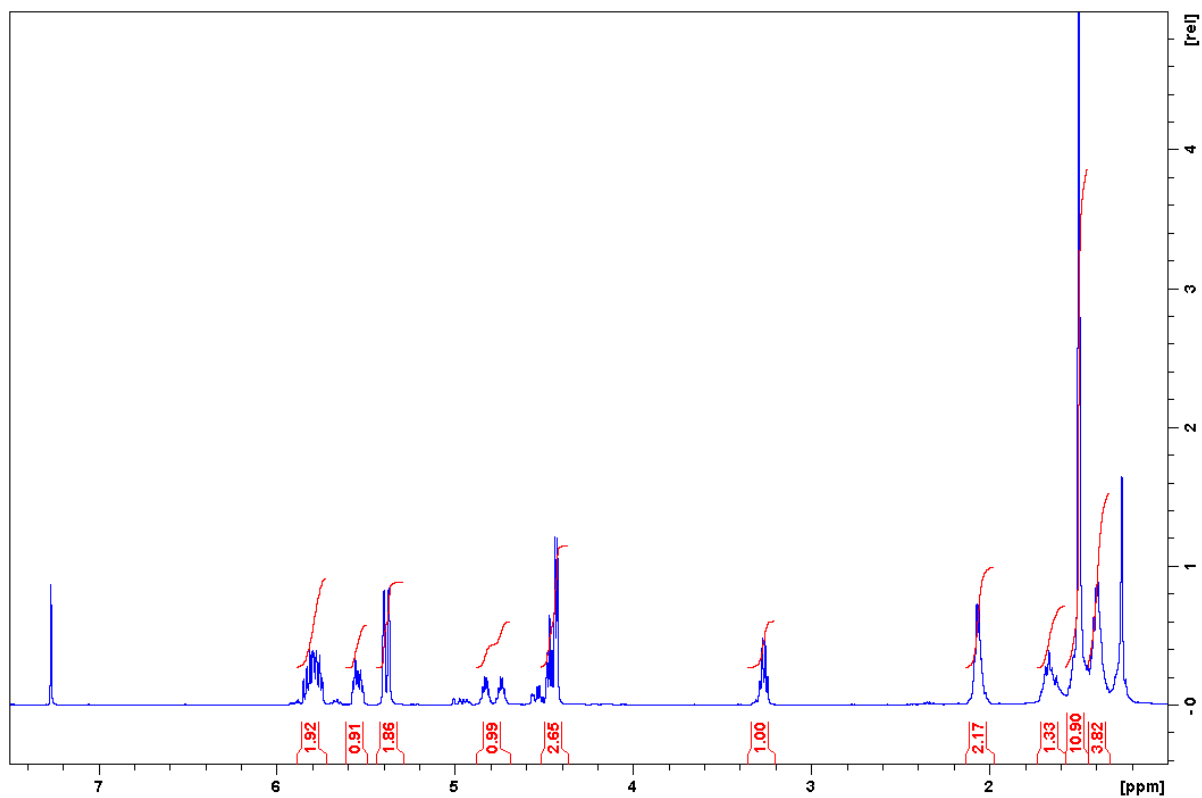
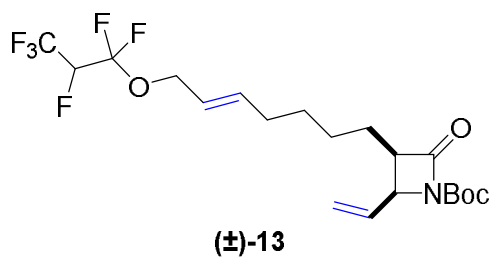


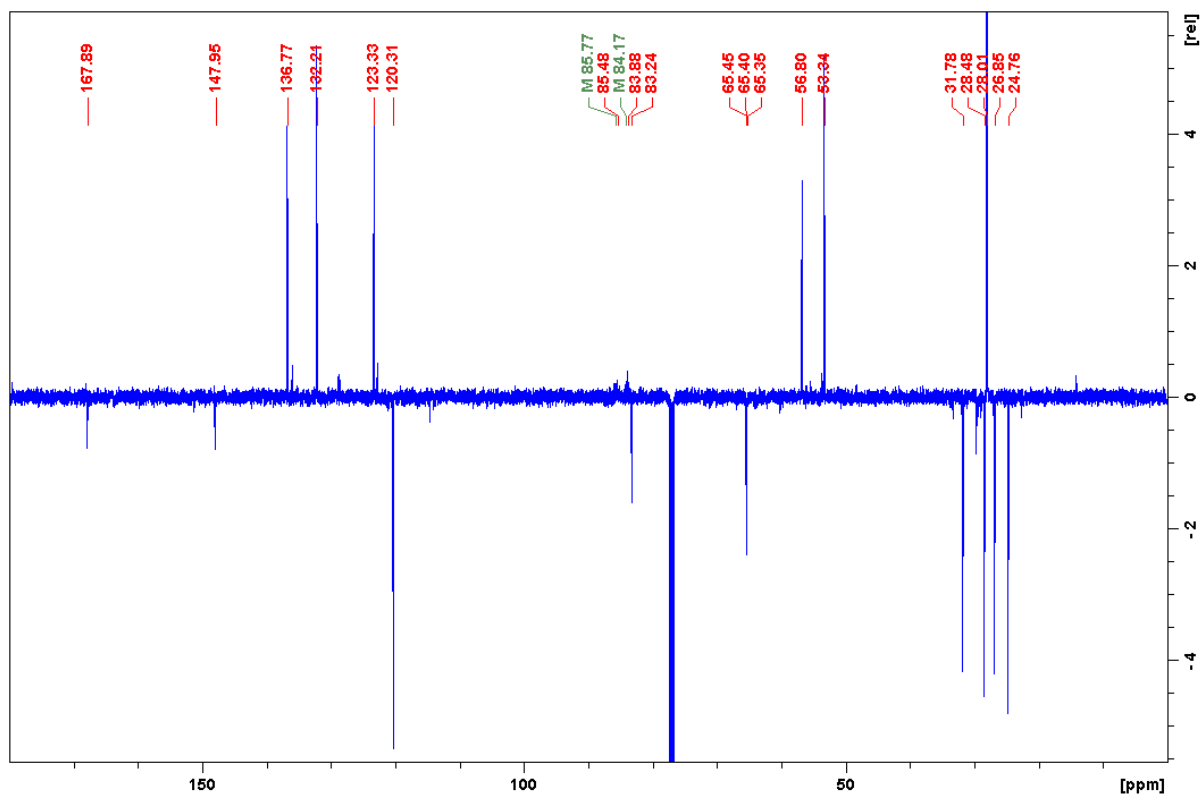
(2*S,3*R**)-*tert*-Butyl 3-((*E*)-7-(1,1,2,3,3,3-hexafluoropropoxy)hept-5-en-1-yl)-2-((*E*)-3-(1,1,2,3,3,3-hexafluoropropoxy)prop-1-en-1-yl)-4-oxoazetidine-1-carboxylate**



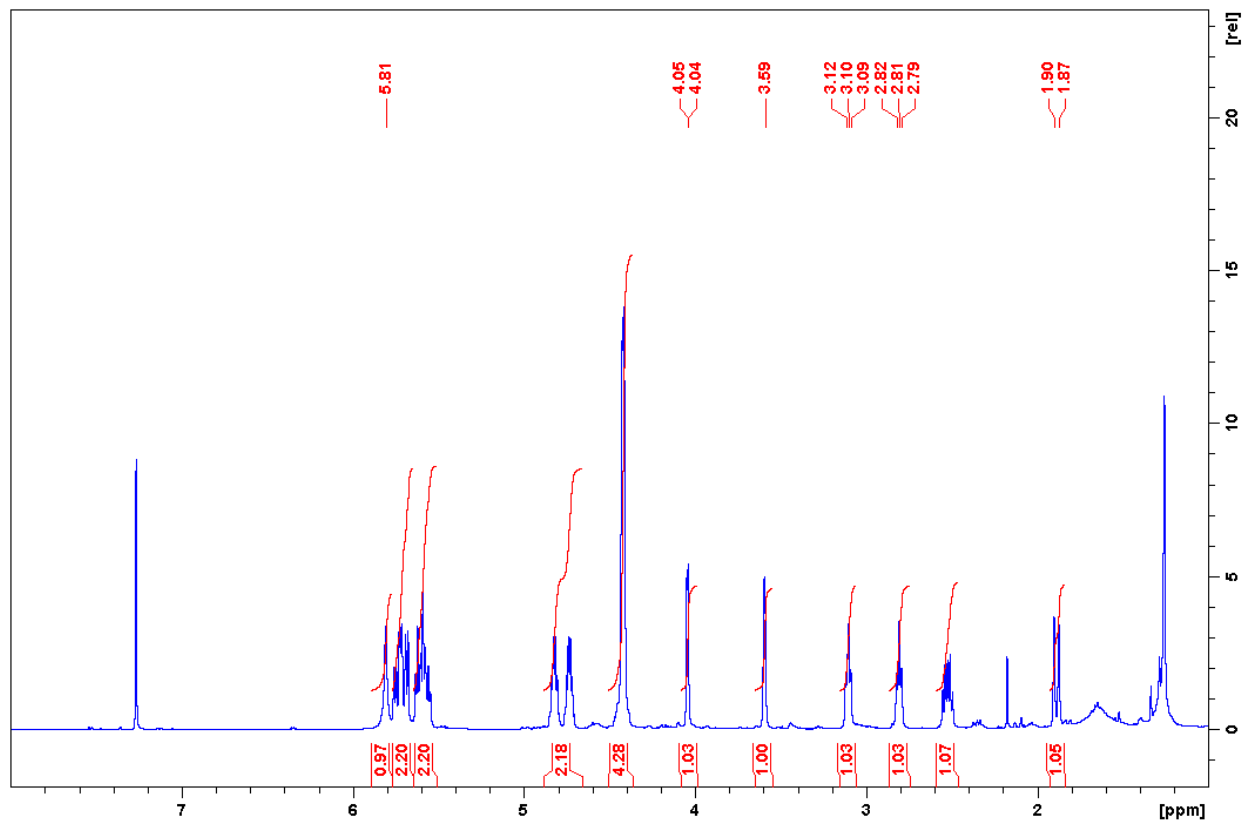
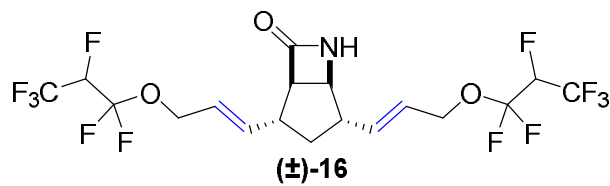


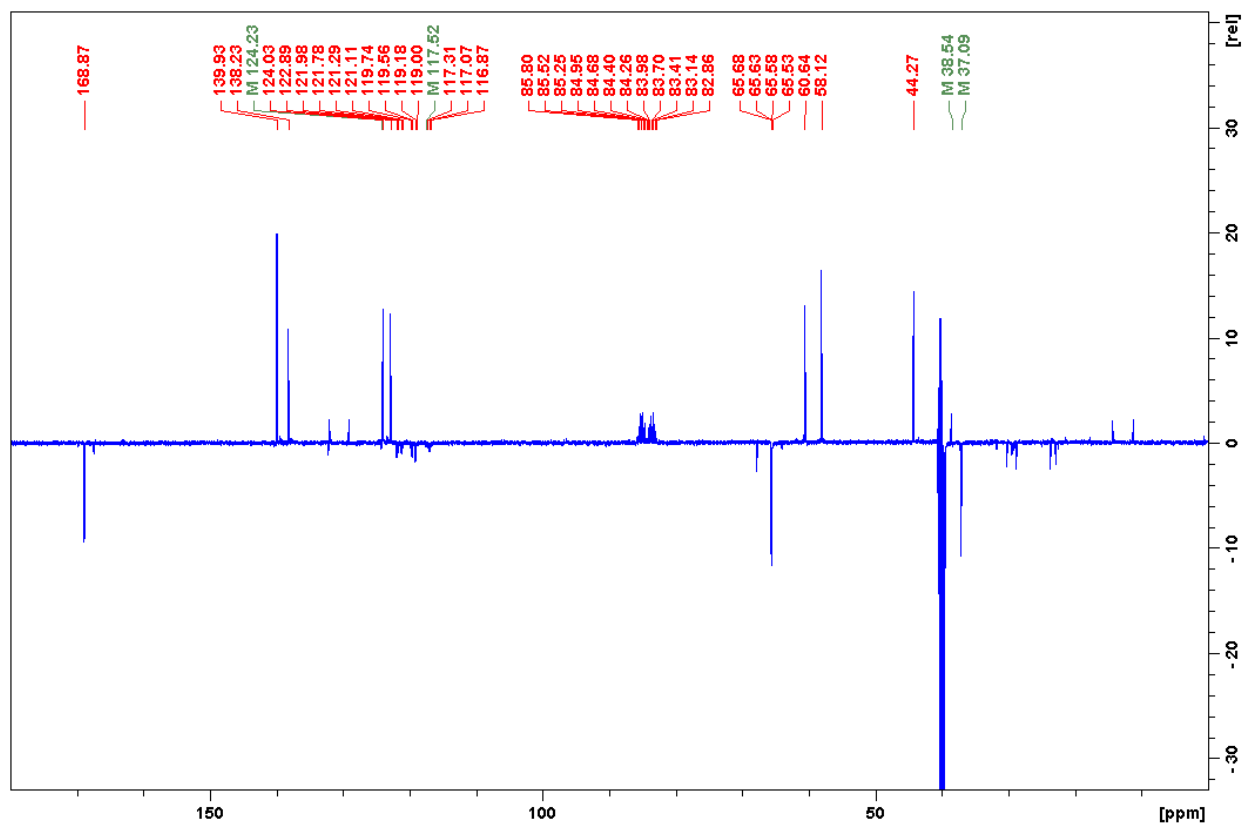
(3*R,4*S**)-tert-Butyl 3-((*E*)-7-(1,1,2,3,3,3-hexafluoropropoxy)hept-5-en-1-yl)-2-oxo-4-vinylazetidine-1-carboxylate**





(1*R,2*R**,4*S**,5*S**)-2,4-bis((*E*)-3-(1,1,2,3,3,3-Hexafluoropropoxy)prop-1-en-1-yl)-6-azabicyclo-[3.2.0]heptan-7-one**





(1*R,2*R**,4*S**,5*S**)-tert-Butyl 2,4-bis((*E*)-3-(1,1,2,3,3,3-hexafluoropropoxy)prop-1-en-1-yl)-7-oxo-6-aza-bicyclo[3.2.0]heptane-6-carboxylate**

